Imaging Core Annual Report FY2018

Dr. Charles Delwiche and Amy Beaven

Table of Contents

Executive Summary	1
Mission of the Facility	2
Introduction and History	2
Organization Structure and Governance	3
Personnel	3
Outreach Activities	4
Summary of Imaging Core Usage	5-8
Operating Cost Analysis	9-12
Proposed Rate Schedules	13
Appendix 1: Past Imaging Core Rates	14
Appendix 2: Summary of Changes in IC Instrumentation	15-16
Appendix 3: Imaging Equipment on Campus	17
Appendix 4: Publications	18-27
Appendix 5: Rates at Other Institutions	28-29

Executive Summary

The Imaging Core (IC) plays a key role in research, training, and education in the life sciences at UMD. In FY18 alone, 56 laboratories from 13 departments in 4 colleges (CMNS, AGNR, ENGR, SPH), plus 4 off-campus users (Biomarker Strategies, Weinberg Medical Physics, the USGS, and Salisbury University) used the facility's high-end light microscopes, as did students in the undergraduate course BSCI421 and the FIRE program. During the past year, the director trained 62 new researchers to independently operate the IC's microscopes, bringing the total number of trained users since FY05 to 799. Access to the facility's microscopes and high-quality training has directly resulted in at least 155 publications (see Appendix 4).

Compared to FY17, when the IC ran a deficit of \$8,734, total income collected via charge-backs from all microscopes exceeded facility expenses by \$30,865 in FY18. This surplus is due to a number of factors, including: 1) the IC spent \$2,420 less on consumables than average, 2) the Advisory Committee directed the IC to purchase a lower-cost (\$9,180 versus \$17,461) service contract on the DeltaVision (a savings of \$8,281), 3) the IC raised hourly rates by \$2 on the confocal and DeltaVision and by \$5 on the AxioObserver and Nikon scopes beginning February 1, 2018 (again, as per the Advisory Committee) and 4) use of the IC's instruments reached historically high levels in FY18 (Figure 1). In fact, FY18 marks the first time the Leica SP5x collected enough income from charge-backs to pay for both the microscope service contract and the (exceedingly expensive) contract on the white light laser.

If rates remain unchanged and microscope use stays at FY18 levels, the IC account balance will slowly increase through FY25 (Figure 6). However, as the instruments continue to age, we predict expenses will increase beyond current average levels due to wear and tear on parts and instruments that are not covered under service contracts, including all the various heavily-used objective lenses and, on the Deltavision, the light source and sCMOS camera, which are not covered under the less expensive service contract. In fact, expenses in FY19 are already trending higher due to the failure of a sensor on the CO2 incubator (>\$1,550 repair).

Looking to the future, the IC's two confocals (purchased in 2008 and 2009) will become obsolete in 2-3 years. Meanwhile, the number of new, high-end microscopes on campus has steadily been increasing, in part due to the IC Director's involvement in the acquisition, management, and installation of the PerkinElmer spinning disk, ASI diSPIM lightsheet, and JPK AFM systems for the newly establish Imaging Incubator. If the IC is to remain competitive, solvent, and self-sustaining, the Director suggests we take the following actions:

- 1. Maintain current rates in FY19, in spite of the surplus in FY18.
- 2. Obtain funding for a new, state-of-the-art, confocal microscope. The Director started this process in May 2017 by submitting an NIH S10 Shared Instrumentation Grant for a Zeiss LSM880 Airyscan. Unfortunately, the grant was not funded, and was therefore resubmitted in May 2018.

Implementing some or all of these recommendations will ensure the IC remains a valuable imaging resource for years to come.

Facility Mission

The mission of the Imaging Core (IC) is to enhance research and education by:

- 1. Providing access to state-of-the-art light microscopy and imaging instrumentation.
- 2. Offering detailed training opportunities and support in basic and advanced light microscopy techniques.
- 3. Keeping researchers up to date with the latest technology and innovations in light microscopy.

Introduction/History

The Imaging Core (IC) was established in the year 2000 by the Department of Cell Biology and Molecular Genetics in order to enhance research and education at the University of Maryland. It provides students and faculty members with access to sophisticated light microscopes and imaging instrumentation whose purchase and maintenance costs far exceed the budgets of individual investigators.

The IC facility is located in 0107 Microbiology building, and includes 10 rooms, five of which are dedicated microscope space, office space for the Director, and a wet-bench lab space with fume hood. When first established, the IC contained a single confocal microscope and a deconvolution microscope. Over the years, demand for time on the instruments increased dramatically, necessitating the purchase of a second confocal in 2008. At present, the IC contains 2 state-of-the-art confocal microscopes (a Zeiss LSM710 and Leica SP5X), a DeltaVision deconvolution/TIRF microscope (installed in March 2014), a Zeiss AxioObserver fluorescence microscope (acquired in February FY14), a Zeiss Axiophot brightfield microscope, and a Nikon Eclipse upright microscope with color camera (transferred to the facility in October 2016).

In December 2014, a PerkinElmer spinning disk confocal microscope was purchased and installed in the Physical Sciences Complex, and became the first instrument in the new Imaging Incubator facility. Following that, an ASI diSPIM was installed in April 2017, and a JPK NanoWizard 4a BioAFM in October 2017. The Director of the IC managed these instruments until transferring all responsibilities to the new Director of the IC, Stephan Brenowitz (hired in July 2017). He is currently assembling a 3-photon system in the new facility.

The Director of the IC, Amy Beaven, oversees the routine operation of the IC and is available during normal business hours to provide training on all equipment, guidance on experimental design, assistance with image analysis, and technician-assisted microscope operation. Since taking over operation of the IC in November 2005, Ms. Beaven has trained over 799 researchers from at least 13 different departments in six colleges and three different campuses (UMCP, UMB, Shady Grove, Salisbury) of the University of Maryland.

The IC is used by a diverse group of investigators, including undergraduates, graduate students, post-docs, technicians and faculty. In addition, students enrolled in the annual 2-credit class CBMG688W/BSCI427, Principles of Microscopy, gained hands-on experience in the operation of the IC's brightfield, DeltaVision and Leica SP5X confocal microscopes until the fall of 2017, when Professor Stephen Wolniak retired. The course trained an average of fifteen students each year for over eleven years.

In the past, funding for the IC came from a combination of user fees and support from the University of Maryland. In an effort to become self-sustaining, trends in facility income, expenses and instrument usage were analyzed over time (the details of which are published in IC's FY10-17 Annual Reports). The analysis showed that a gradual increase in hourly instrument rates were necessary in order for the facility to become financially independent. As such, user fees were incrementally increased over several years in the hopes that the IC would be able to cover all maintenance and service contract costs through user fees alone. It should be noted that current instrument fees are priced competitively and still below the average rates charged at similar institutions with equivalent instrumentation (see Appendix 5).

Please see Appendix 3 for a complete list of current instrumentation, and Appendix 2 for changes in IC instrumentation since FY05.

Organizational Structure and Governance

- Director of the Facility: Amy Beaven
- Faculty supervisor: Dr. Charles Delwiche, Professor
- Advisory Committee: Charles Delwiche, Professor (CBMG), Jose Feijo, Professor (CBMG), Iqbal Hamza, Professor (ANSC), Wolfang Losert, Professor (PHYS/IPST), Giuliano Scarcelli, Assistant Professor (ENGR), Arpita Updadhyaya, Associate Professor (PHYS), Patrick Kanold, Professor (Biology), and Stephan Brenowitz (Director, Imaging Incubator)

Personnel

The Director of the Facility, Amy Beaven, is the only full-time staff member within the facility. She was hired in 2005 to manage the Imaging and Genomics Core facilities and was promoted to Director in 2010. Ms. Beaven received her Master's degree in Biology in 1999 and has over 14 years of intensive experience in imaging techniques, including laser scanning confocal, spinning disk confocal, deconvolution, and TIRF. She is available during the hours of 8am-4:30pm to provide guidance in experimental design, training on all equipment, technician-assisted confocal operation and assistance with image analysis.

Outreach Activities During FY18

- 1. July 2017: Dr. Stephan Brenowitz was hired as Director of Imaging Incubator on July 1. Over the next few months, the director of the IC assisted with operation, training, and billing of instruments (PerkinElmer spinning disk, ASI diSPIM) until all responsibilities were handed over to Stephan in October.
- 2. July August 2017: the Director supervised federal-work-study student Hawa Anthony, who assisted with billing and the preparation of the FY17 Annual Report.
- 3. August 2017: the Director continued to assist BioENGR/IBBR with their Zeiss LSM 700 system by training students to use the scope.
- 4. July November, 2017: The director of the IC designed a new website for the Bioscience Core Facilities (Proteomics, Flow, Genomics, Imaging Core, Imaging Incubator). The site was officially launched on 11/9/2017. http://biosciencecores.umd.edu/
- 5. October 2017: The IC's old 1999 Deltavision scope (Nikon TE200) was transferred to the PSC Imaging Incubator so that it could be incorporated into a future AFM.
- 6. October 2017: The IC director designed and presented Bioscience Core Facilities poster at Bioscience Day.
- 7. October 16-18: The IC director oversaw installation of JPK NanoWizard 4a AFM microscope in the Imaging Incubator.
- 8. January 2017: The IC hosted the demo of a BioTek Lionheart fluorescence microscope.
- 9. February 9-16, 2018: The IC hosted and coordinated the demo of a Zeiss 800 Airyscan in order to collect additional preliminary data for a grant proposal.
- 10. March 2018: The IC director assisted members of the Space Biofilm project to take images of the samples they sent to the International Space Station via SpaceX CRS-12 (SpaceX-12: Falcon 9 rocket, Dragon spacecraft). https://www.launch.umd.edu/project/5877
- 11. March 2018: The IC director submitted an on-campus limited submission preproposal for a Zeiss LSM 880 Airyscan Fast confocal (resubmission of last year's proposal).
- 12. March May 2018: the Director trained Teaching Assistants in BSCI421 (Cell Biology) to use Zeiss AxioObserver and Nikon Eclipse so that they could assist undergraduate students to take images for their laboratory experiments.
- 13. May 3, 2018: Imaging Incubator Grand opening. The director of the IC participated by providing tours of the facility.
- 1. May 31, 2018: The Director coordinated with NIH-funded faculty in CBMG, Biology, ANSC, and Physics to resubmit the full NIH S10 Shared Instrumentation Grant for the purchase of a new Zeiss LSM880 Airyscan Fast (proposal under review).
- 14. June 2018: The IC director trained 10 FIRE undergraduate students to use the AxioObserver for their research projects.

Summary of Facility Usage

This year, in order to collect more accurate instrument usage data, the director began tracking both the billable hours for each instrument, as well as the actual hours each instrument was scheduled/booked and therefore unavailable to other users (Table 1).

In FY18, the Zeiss LSM710 was scheduled an average 23 hours/week and Leica SP5X was used 33 hours/week, though use of the instrument increased to \sim 48 hours/week in the February-May 2018 time-frame. The combined average usage of 56 hours/week is a 29% increase from FY17.

From FY09 to FY14, the Zeiss consistently saw more use than the Leica, but the trend reversed in FY15. Over the past 4 years, the Leica was used on average 397 more hours per year than the Zeiss. This is most likely because the Leica was upgraded with a new, more sensitive detector (HyD detector and time-gating technology) in FY14.

The Deltavision was used an average of 8 hours per week—a 44% increase over FY17. Use of the AxioObserver dropped 93 percent, but Nikon Eclipse use increased 139%.

Table 1: Leica SP5X Summary Data:

Fiscal Year	Income	Total # Hours Charged	Total # Hours Booked	Total Hrs used for UMCP Courses/Programs	Total # Training Sessions
2009	\$5,091	346	N/A	0	39
2010	\$18,363	1283	N/A	71	43
2011	\$24,290	1325	N/A	56	35
2012	\$21,882	1021	N/A	63	29
2013	\$21,922	932	N/A	91	35
2014	\$25,161	886	N/A	71	34
2015	\$25,576	1206	N/A	39	27
2016	\$35,865	1602	N/A	36	17
2017	\$31,802	1153	N/A	6	20
2018	\$49,599	1569	1720	0	28
Total	\$259,552	11323	N/A	433	307

Table 2: Zeiss LSM710 Summary Data:

Fiscal Year	Income	Total # Hours Charged	Total # Hours Booked	Total Hrs used for UMCP Courses/Programs	Total # Training Sessions
2010	\$12,370	804	N/A	0	44
2011	\$33,448	1762	N/A	0	33
2012	\$27,895	1244	N/A	0	20
2013	\$33,889	1248	N/A	0	26
2014	\$31,471	1174	N/A	1.5	17
2015	\$27,655	964	N/A	6.5	22
2016	\$32,847	1021	N/A	7	20
2017	\$26,055	688	N/A	29	30
2018	\$35,400	1091	1192	0	13
Total	\$261,030	9996	N/A	44	225

Table 3: DeltaVision Summary Data:

Fiscal Year	Income	Total # Hours Charged	Total # Hours Booked	Total Hrs used for UMCP Courses/Programs	Total # Training Sessions
2014	\$317	32	N/A	0	12
2015	\$6472	376	N/A	24	13
2016	\$6,824	270	N/A	33	20
2017	\$7267	270	N/A	20	21
2018	\$12,674	423	486	0	6
Total	\$33,554	1371	N/A	77	72

Table 4: AxioObserver Fluorescence Microscope Data

Fiscal Year	Income	Total # Hours Charged	Total # Hours Booked	Total Hrs used for UMCP Courses/Programs	Total # Training Sessions
2014	\$113	23	N/A	0	0
2015	\$790	155	N/A	0	1
2016	\$1,086	198	N/A	0	5
2017	\$2066	319	N/A	0	22
2018	\$975	117	130	36	8
Total	\$5,030	812	N/A	36	36

Table 5: Nikon Eclipse Microscope Data

Fiscal Year	Income	Total # Hours Charged	Total # Hours Booked	Total Hrs used for UMCP Courses/Programs	Total # Training Sessions
2017	\$103	16	N/A	0	1
2018	\$1025	90	101	7	7
Total	\$1128	106	N/A	7	8

Table 5: Combined Microscope Data by Fiscal Year:

Fiscal Year	Income	Total # Hours Charged	Total # Hours Booked	Total Hrs used for UMCP Courses/Programs	Total # Training Sessions
2009	\$5,091	346	N/A	0	39
2010	\$30,733	2,086	N/A	71	87
2011	\$57,739	3,088	N/A	56	68
2012	\$49,778	2,265	N/A	63	49
2013	\$55,811	2,180	N/A	91	61
2014	\$57,061	2,114	N/A	73	63
2015	\$60,175	2,700	N/A	70	63
2016	\$76,622	3,090	N/A	76	62
2017	\$67,194	2,636	N/A	83	93
2018	\$100,126	3290	3630	36	62
Total	\$560,330	3290	N/A	619	647*

^{*}Including the old Zeiss LSM510, the total # training sessions increases to 799

During FY18, 56 laboratories from 13 departments in 4 colleges (CMNS, AGNR, ENGR, SPH), plus 4 off-campus users (two companies, the USGS, and Salisbury University) made use of the facility's confocal microscopes (Figures 2-3). CBMG accounted for 58% of the total microscope use (Figure 4). Departments within CMNS accounted for 83% of use (Figure 5).

Figure 1: Total number of hours used on Imaging Core instruments by fiscal year

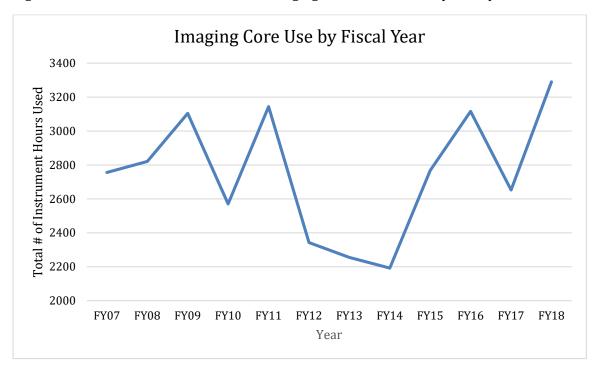


Figure 2: Top Microscope Users by Department FY18

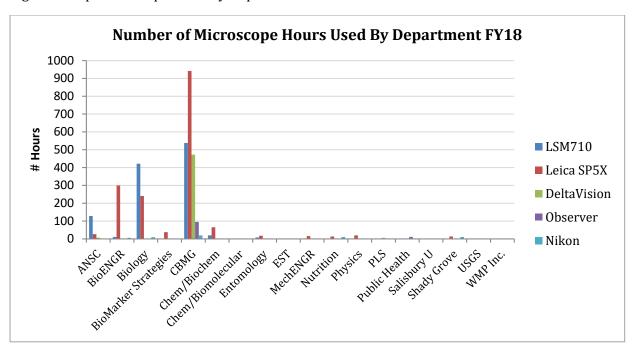


Figure 3: Top Microscope Users FY18

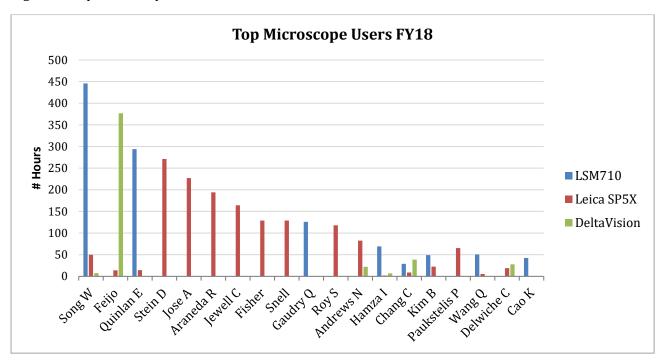


Figure 4: CBMG versus non-CBMG Use of Microscopes FY18

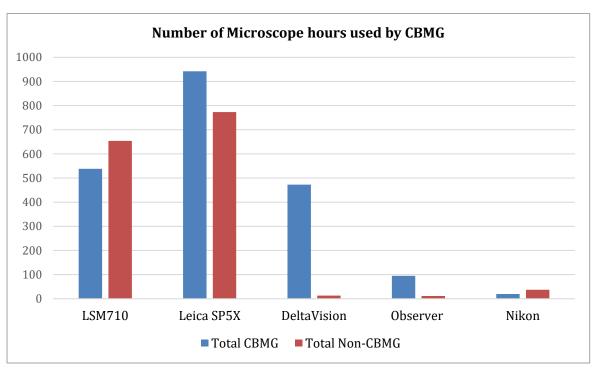
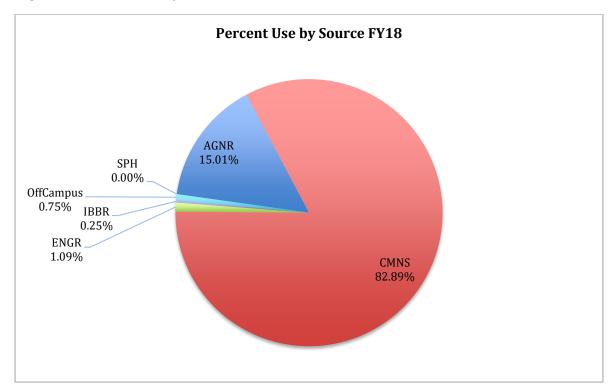


Figure 5: Percent Use by Source FY18



Operating Cost Analysis

Compared to FY17, when the IC ran a deficit of \$8,734, total income collected via charge-backs from all microscopes in exceeded facility expenses by \$30,865 in FY18. This surplus is due to a number of factors, including 1) lower than average consumable expenses in FY18, 2) purchase of a lower-cost (\$9,180 versus \$17,461 in FY17) service contract on the DeltaVision, 3) a \$2 increase in hourly confocal and DeltaVision rates and a \$5 increase on the AxioObserver and Nikon scopes (changes implemented on February 1, 2018), and 4) use of the IC's instruments in FY18 reached historically high levels (Figure 1).

Table 6: FY18 Income and Expenses by Microscope

Instrument/ Source Income	Service contract cost	Consumable Cost	Total Expenses	Income	Income - Expenses
Zeiss LSM 710	\$19,969	\$96	\$20,066	\$35,400	\$15,334
Leica SP5 X	\$39,630	\$96	\$39,726	\$49,600	\$9,873
DeltaVision	\$9,180	\$96	\$9,276	\$12,674	\$3,397
Zeiss Observer	\$0	\$96	\$96	\$975	\$879
Axiophot	\$0	\$0	\$0	\$3	\$3
Nikon Eclipse	\$0	\$96	\$96	\$1,025	\$929
LSM700 IBBR	N/A	N/A	N/A	\$450	\$450
Total	\$68,779	\$482	\$69,261	\$100,126	\$30,865

Table 7: Total Imaging Core Facility Income and Expenses from FY09-FY18

Year	Total Income from Charge-backs*	Total Imaging Core Expenses	Net Balance*
FY2009	\$5,091	\$6,113	(\$1,023)
FY2010	\$30,732	\$29,564	\$1,169
FY2011	\$57,738	\$55,525	\$2,214
FY2012	\$49,778	\$76,562	(\$26,685)
FY2013	\$55,810	\$59,673	(\$3,862)
FY2014	\$57,184	\$59,876	(\$2,692)
FY2015	\$60,303	\$73,955	(\$13,652)
FY2016	\$79,028	\$75,544	\$3,484
FY2017	\$71,458	\$80,234	(\$8,784)
FY2018	\$100,126	\$69,261	\$30,864
Total	\$567,250	\$586,315	(\$19,065)

^{*} If instrument income from subsidies associated with the Leica SP5x NSF MRI grant (\$37,500 a year in FY10, FY11, and FY12) is included, the net balance for all instrument at the end of FY18 is \$93,435. The actual account balance in June of 2018 was \$112,863.

Table 8: Cost Breakdown: Leica SP5X

Year	Service Contract Cost	Expenses	Income	Income (subsidies)	Income - Expenses
FY2009	0	\$6,113	\$5,091	0	(\$1,023)
FY2010	\$26,000	\$2,376	\$18,363	\$37,500	\$27,487
FY2011	\$36,075	\$488	\$24,290	\$37,500	\$25,227
FY2012	\$52,296	\$2,056	\$21,882	\$37,500	\$5,032
FY2013	\$37,092	\$1,887	\$21,922	0	(\$17,057)
FY2014	\$37,092	\$360	\$25,160	0	(\$12,290)
FY2015	\$37,743	\$51	\$25,576	0	(\$12,218)
FY2016	\$37,743	\$609	\$35,865	0	(\$2,487)
FY2017	\$39,630	\$616	\$33,158	0	(\$7,088)
FY2018	\$39,630	\$96	\$49,600	0	\$9,873
Total	\$303,670	\$15,212	\$211,308	\$112,500	\$13,443

Table 9: Cost Breakdown: LSM 710

Year	Service Contract Cost	Expenses	Income	Income (subsidies)	Income - Expenses
FY2009	0	0	0	0	0
FY2010	0	\$1,188	\$12,370	0	\$11,182
FY2011	\$17,730	\$1,232	\$33,448	0	\$14,487
FY2012	\$19,260	\$2,951	\$27,896	0	\$5,685
FY2013	\$19,260	\$1434	\$33,889	0	\$13,195
FY2014	\$19,260	\$360	\$31,471	0	\$12,211
FY2015	\$19,755	\$51	\$27,655	0	\$7,848
FY2016	\$19,755	\$608	\$32,847	0	\$12,484
FY2017	\$19,969	\$785	\$27,080	0	\$6,326
FY2018	\$19,969	\$96	\$35,400	0	\$15,334
Total	\$134,989	\$8,608	\$226,655	0	\$83,418

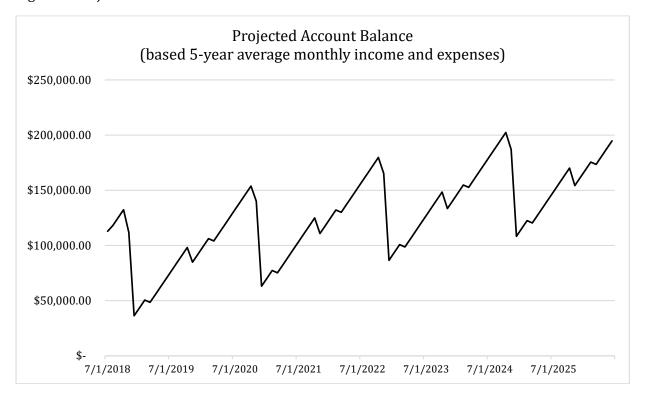
Table 10: Cost Breakdown: DeltaVision

Year	Service Contract Cost	Expenses	Income	Income (subsidies)	Income - Expenses
FY2014	\$0	\$739	\$371	0	(\$422)
FY2015	\$16,200	\$51	\$6135	0	(\$10,097)
FY2016	\$16,200	\$323	\$6,824	0	(\$9,700)
FY2017	\$17,461	\$160	\$8,623	0	(\$8,998)
FY2018	\$9,180	\$96	\$12,674	0	\$3,397
Total	\$49,861	\$1,273	\$21,919	0	(\$29,216)

Projected Cost Analysis:

If rates remain unchanged and microscope use stays at FY18 levels, the IC account balance will continue to grow past 2025 (Figure 6). This projection does not include potential large one-time expenses, such as objective lens replacements (~\$2000 - \$10,000 depending on the lens), and parts no longer covered under the DeltaVision's more limited service contract (light source and sCMOS camera). FY19 expenses are already trending above average due to the failure of a sensor on the CO2 incubator (>\$1,550 repair).

Figure 6: Projected Account Balance



Proposed Rate Schedule:

A thorough examination of confocal microscope rates at other institutions (Appendix 5) shows that the facility's confocal peak rates are priced slightly lower than the average hourly rate of \$36, and the DeltaVision's rates are on par with the average. The IC continues to offer highly competitive, low-priced off-peak rates of \$24/hr. The director suggests keeping these rates the same in FY19. In addition, after speaking with the Sponsored Programs Accounting & Compliance office, the director suggests we eliminate the rate for "UM users not affiliated with CMNS, AGNR, and ENGR" category, as it does not comply with UM rate guidelines for sponsored programs.

Table 11: Suggested Imaging Core Rates for Academic Year 2019

Instrument	Users within CMNS, AGNR (excluding Vet Med), ENGR	Eliminate: UM users not affiliated with CMNS, AGNR, ENGR	Users not affiliated with campus
Leica SP5X, Zeiss LSM710, PerkinElmer Spinning Disk, Hourly Rate for Unassisted Use	\$34/hr peak \$24/off-peak	\$45/hr	\$85/hr
DeltaVision Hourly Rate for Unassisted Use	\$30/hr peak \$24/hr off-peak	\$45/hr	\$85/hr
Leica SP5X, Zeiss LSM710, DeltaVision, PerkinElmer Spinning Disk 14 hour overnight flat fee, 6pm-8am, M-F	\$150	Inquire	Inquire
Leica SP5X, Zeiss LSM710, DeltaVision, PerkinElmer Spinning Disk 12 hour day rate (flat fee), 8am-8pm, M-F	\$250	Inquire	Inquire
Leica SP5X, Zeiss LSM710, DeltaVision, PerkinElmer Spinning Disk 24 hour rate (flat fee), M-F	\$350	Inquire	Inquire
Leica SP5X, Zeiss LSM710, DeltaVision, PerkinElmer Spinning Disk 24 hour rate (flat fee), Weekend Rate	\$200	Inquire	Inquire
Leica SP5X, Zeiss LSM710, DeltaVision, PerkinElmer Spinning Disk 48 hour rate (flat fee), Weekend Rate	\$400	Inquire	Inquire
Leica SP5 X, Zeiss LSM 710, DeltaVision, PerkinElmer Spinning Disk Hourly Rate for Assisted Use	\$50/hr	\$60/hr	\$180/hr
Leica SP5X, Zeiss LSM 710, DeltaVision, PE Training Fee (per person)	\$150	\$200	\$300
Zeiss AxioObserver, Axiophot and Nikon Eclipse Hourly Rate for Unassisted Use	\$10/hr	\$20/hr	\$40/hr
Zeiss AxioObserver, Axiophot and Nikon Eclipse Training Fee (per person)	\$25	\$40	\$100

Appendix 1: Past Imaging Core Rates

Equipment	Location	Purchase and/or Installation Date	In-College Rate History (Academic Year)
Zeiss LSM 710 Confocal	0107E MICB	October 2009 (Purchased by	2009/2010: \$15.00/hr 2010/2011: \$18.60/hr
Microscope		CBMG/CMNS)	2011/2012: \$22.00/hr 2012/2013: \$26.00/hr Peak; \$23.00/hr Off-Peak 2013/2014: \$26.00/hr Peak; \$23.00/hr Off-Peak
			2014/2015: \$28.00/hr Peak; \$24.00/hr Off-Peak 2015/2016: \$32.00/hr Peak; \$22.00/hr Off-Peak 2016/2017: \$32.00/hr Peak; \$22.00/hr Off-Peak
Lain CDEV	010711	D	2/1/2018: \$34.00/hr Peak; \$24.00/hr Off-Peak
Leica SP5X Confocal	0107H MICB	December 2008 (Purchased via NSF	2008/2009: \$15.00/hr 2009/2010: \$15.75/hr
Microscope		MRI grant)	2010/2011: \$18.60/hr
			2011/2012: \$22.00/hr
			2012/2013: \$26.00/hr Peak; \$23.00/hr Off-Peak
			2013/2014: \$26.00/hr Peak; \$23.00/hr Off-Peak 2014/2015: \$28.00/hr Peak; \$24.00/hr Off-Peak
			2015/2016: \$32.00/hr Peak; \$22.00/hr Off-Peak
			2016/2017: \$32.00/hr Peak; \$22.00/hr Off-Peak
			2/1/2018: \$34.00/hr Peak; \$24.00/hr Off-Peak
Deltavision	0107F	March 2014	2013/2014: \$10/hr
Deconvolution/	MICB	(Purchased by	2014/2015: \$10/hr thru 3/2015
TIRF Microscope		CBMG)	2014/2015: \$28.00/hr Peak; \$24.00/hr Off-Peak 2015/2016: \$28.00/hr Peak; \$22.00/hr Off-Peak
			2016/2017: \$28.00/hr Peak; \$22.00/hr Off-Peak
			2017/2018: \$30.00/hr Peak; \$24.00/hr Off-Peak
Zeiss	0107K	February 2014	2013 to 2017: \$5/hr
Axio0bserver		(Relocated from	2/1/18: \$10/hr
Fluorescence		BRB; originally	
		purchased by CBMG)	
Axiophot	Main lab	CoolSnap, Elements:	\$2.00/hr since installation
Fluorescence		July 2007	,
Microscope		(purchased by	
Nilson Ealings	01070	CBMG) October 2017	2016 /2017, ¢f /bm
Nikon Eclipse	0107B	(relocated)	2016/2017: \$5/hr 2/1/18: \$10/hr
PerkinElmer	B0118	December 2014	2014/2015: \$28.00/hr Peak; \$24.00/hr Off-Peak
confocal spinning	PSC	(purchased by	2015/2016: \$32.00/hr Peak; \$22.00/hr Off-Peak
disk		Physics for PSC)	2016/2017: \$32.00/hr Peak; \$22.00/hr Off-Peak
			2/1/18: \$34.00/hr Peak; \$24.00/hr Off-Peak

Appendix 2: Summary of changes in IC instrumentation since November 2005

- 8/2006: A Mini Med 90 Film Processor (cost: \$3,588.00) replaced the old Konica processor. The department paid \$2,500.00 of the total cost and each of the following PIs contributed \$109: Jonathan Dinman, Jeffrey DeStefano, Kenneth Frauwirth, David Mosser, Anne Simon, Wenxia Song, Richard Stewart and Elizabeth Gantt.
- 10/2006: Dr. Steve Wolniak (Interim Chair of CBMG) procured a Zeiss Axiophot fluorescence microscope for the facility following Dr. Ron Weiner's retirement. A CoolSnap EZ monochrome camera, computer workstation and Nikon Elements software (total cost: \$13,400.00) were purchased in 2007 for the microscope using CBMG funds.
- 4/2007: The 7700 Sequencer Detector was replaced with a Roche LightCycler 480 Real-Time PCR machine, which was purchased by CBMG using the Bioscience Research Building capital equipment funds (and is housed in BRB; see below).
- 8/2007: Due to a drop in usage, the 3100 North DNA sequencer was taken out of operation.
- 12/2008: The instruments in 0107H MICB (two ABI 3100 DNA sequencers, the ABI 3730xl DNA Sequencer and the Roche LightCycler 480 Real-Time PCR machines) were moved to the new Genomics Core, room 2229 Bioscience Research Building.
- 12/2008: The Leica SP5 X confocal microscope was installed in room 0107H MICB. This microscope was obtained by Drs. Ian Mather and. Steve Wolniak via an NSF MRI grant.
- 10/2009: The LSM510 confocal microscope was dismantled to make way for the new Zeiss LSM710, which was purchased using college funds, authorized by Dean Allewell.
- 1/2009: Genomics Core Equipment: Bio-Rad CFX 96 Real-time PCR machine was purchased and placed in room 2229 BRB.
- 4/2010: Genomics Core Equipment: Due to a drop in usage, the 3100 "West" DNA sequencer was taken out of operation.
- 7/2011: Genomics Core Equipment: July 2011: Both the 3100 "West" and 3100 "North" DNA sequencers were sold through Terrapin Trader.
- 11/2011: A Thermo Scientific Midi 40 CO2 incubator was purchased using IC funds (\$3,194.00).
- 2/2014: a Zeiss AxioObserver widefield fluorescence microscope was relocated from room 3207 Bioscience Research Building. The microscope was installed in room 0107K and upgraded with a new computer, new software (Zen 2012) and a new power supply, using a combination of departmental and IC funds.
- 3/2014: A DeltaVision Deconvolution/TIRF microscope was installed in room 0107F MICB. The microscope was purchased with departmental and college funds.
- 4/2014: Financial responsibility for the DNA sequencer was transferred to the Biology Dept.
- 8/2014: The Leica SP5 X was upgraded with a new HyD detector and time-gating technology.
- 10/2014: The Zeiss LSM 710 computer was replaced (free upgrade due to computer issues with the old operating system) with a Windows 7 computer and the software was upgraded to the latest version of Zen.
- 12/2014: A PerkinElmer spinning disk confocal microscope was installed in the Satellite Core, room B0118 Physical Sciences Complex.
- 1/2015: As a result of increased Imaging Core responsibilities, management of the Genomics Core was transferred from Amy Beaven to Dr. Yan Wang.
- 7/24/15: The Mini Med 90 Film Processor was taken out of service. It was decontaminated by DES (Cleveland Taylor) and sent to Terrapin Trader.
- 2/2016: The IC implemented a new BSL-2 protocol to allow imaging of approved BSL-2 samples.

- 4/2016: Dr. Charles Delwiche purchased 4 new objective lenses for the DeltaVision and loaned them to the IC: 10x 0.4 NA, 20x 0.75NA, 40x 0.6NA LWD and 40x 1.3NA oil lens.
- 10/2016: A Nikon Eclipse transferred from Dr. Nicole Li's old lab on the 3rd floor of MICB to the IC, room 0107A MICB.
- 4/2017: the ASI diSPIM lighsheet microscope was installed in B0118 PSC on April 4-5 (ahead of the purchase order issued on April 27, 2017).
- 5/2017: The IC acquired room 0105 MICB. The LightCycler 480 qPCR machine was transferred from 2229 BRB to this space, and the Director of the IC now manages the instrument (again).
- 10/7/17: Old DeltaVision Nikon microscope moved to Imaging Incubator in PSC. To be incorporated in the JPK NanoWizard 4a BioAFM.
- 10/16 10/18/2017: JPK NanoWizard 4a BioAFM installed (purchase order issued 8/18/17).
- 11/9/2017: Launched new website: http://biosciencecores.umd.edu/

Appendix 3: Imaging Equipment on Campus

	Purchase/	Owner		Available to
Equipment	Installation	(purchased	Location	anyone in
	Date	by)		CBMG?
Zeiss LSM 710	10/2009	CBMG/CMNS	CBMG Imaging Core:	Yes
Confocal			0107E MICB	
Microscope	12/2000	ANICC /CDMC	CDMC Inventor Come	Yes
Leica SP5X Confocal	12/2008	ANSC/CBMG (Purchased via	CBMG Imaging Core: 0107H MICB	res
Microscope		NSF MRI grant)	010711 MICD	
Deltavision	3/2014	CBMG	CBMG Imaging Core:	Yes
Deconvolution/			0107F MICB	
TIRF Microscope				
Zeiss	2/2014	CBMG	CBMG Imaging Core:	Yes
AxioObserver		(Relocated from	0107K	
Fluorescence	_ /0.0.	BRB in 2014)		
Axiophot	7/2007	CBMG	CBMG Imaging Core:	Yes
Brightfield Microscope		(Camera and software)	Main lab	
Nikon Eclipse	10/2017	CBMG	CBMG Imaging Core:	Yes
w/color camera	10/2017	(relocated	0107B	163
Wy color camera		fromLi lab)	010715	
PerkinElmer	12/2014	CMNS	CMNS Imaging	Yes
confocal	,	(purchased with	Incubator:	
spinning disk		PSC building	B0118 PSC	
		funds)		
ASI diSPIM	4/2017	CMNS	CMNS Imaging	Yes
Lighsheet		(purchased with PSC building	Incubator: B0118 PSC	
		funds)	DUITOFSC	
JPK NanoWizard	10/2017	CMNS	CMNS Imaging	Yes
4a BioAFM		(purchased with	Incubator:	
		PSC building	B0118 PSC	
		funds)		
Andor Spinning	August 2016	Sougata Roy	Roy Lab, BRB	No
Disk	. l 0040	(CBMG)	E404A PLG1 IIII	
Zeiss LSM700	~July 2012	IBBR/BioENGR	5131A PLS building	No
Confocal Zeiss LSM700	Transferred	Margaret Scull	Scull lab, BRB	No
Confocal	in FY17 from	(CBMG)	Jeun lau, DND	110
domocar	S. Wolniakd	(dDi-id)		
Zeiss LSM800	FY16	Lisa Taneyhill	Taneyhill lab, ANSC	No
Airyscan		(ANSC)		
Confocal				
Zeiss LSM710	~FY14	Murphy/Peer	Murphy/Peer lab, PLS	No
Confocal	FILLOC	(PLS)	A 1 11	
PerkinElmer	FY08	Norma Andrews	Andrews lab, BRB	No
Spinning Disk		(CBMG)		

Appendix 4: Publications

Publications entailed the use of the Zeiss LSM 510:

- 1. Bish, S. E., W. Song, and D.C. Stein. 2008. Quantification of bacterial invasion into host cells using a beta-lactamase reporter strain: Neisseria gonorrhoeae invasion into cervical epithelial cells requires bacterial viability. Microbes Infect. 10:1182-1191.
- 2. Sikes, J. M. & Bely, A. E. Radical modification of the A-P axis and the evolution of asexual reproduction in Convolutriloba acoels. Evolution and Development 10, 619-631 (2008).
- 3. The MHC class II-associated invariant chain interacts with the neonatal Fc gamma receptor and modulates its trafficking to endosomal/lysosomal compartments. Ye L, Liu X, Rout SN, Li Z, Yan Y, Lu L, Kamala T, Nanda NK, Song W, Samal SK, Zhu X. J Immunol. 2008 Aug 15;181(4):2572-85
- 4. Activation of the JAK/STAT-1 signaling pathway by IFN-gamma can down-regulate functional expression of the MHC class I-related neonatal Fc receptor for IgG. Liu X, Ye L, Bai Y, Mojidi H, Simister NE, Zhu X. J Immunol. 2008 Jul 1;181(1):449-63.
- 5. Identification and characterization of an alternatively spliced variant of the MHC class I-related porcine neonatal Fc receptor for IgG.
- 6. Ye L, Tuo W, Liu X, Simister NE, Zhu X. Dev Comp Immunol. 2008;32(8):966-79. NF-kappaB signaling regulates functional expression of the MHC class I-related neonatal Fc receptor for IgG via intronic binding sequences. Liu X, Ye L, Christianson GJ, Yang JQ, Roopenian DC, Zhu X. J Immunol. 2007 Sep 1;179(5):2999-3011
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- 8. Onabajo, O., M. Seeley, A. Kale, B. Qualmann, M. Kessels, S-H. Tan, and W. Song. 2008. Mammalian actin-binding protein 1 regulates BCR-mediated antigen processing and presentation in response to BCR activation. J. Immunol. 180(10):6685-95.
- 9. Sharma, S., Orlowski G. and W. Song. 2009. Btk regulates BCR-mediated antigen processing and presentation by controlling the actin cytoskeleton dynamics in B cells. J. Immunol. 182: 329–339.
- 10. Dong CH, Rivarola M, Resnick JS, Maggin BD and Chang C (2008) Subcellular co-localization of Arabidopsis RTE1 and ETR1 supports a regulatory role for RTE1 in ETR1 ethylene signaling. Plant Journal 53(2): 275-286
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- 13. Song, W., L. Ma, R. Chen, and D. C. Stein. 2000. Role of lipooligosaccharide in Opa-independent invasion of Neisseria gonorrhoeae into human epithelial cells. J. Exp. Med. 191 (6):949-60.
- 14. Cheng, P. C., B. K. Brown, W. Song, and S. K. Pierce. 2001. Translocation of the B cell antigen receptor into lipid rafts reveals a novel step in signaling. J. Immunol. 166 (6):3693-701.
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- 16. Brown, B. K., and W. Song. 2001. The actin cytoskeleton is required for the trafficking of the B cell antigen receptor to the late endosomes. Traffic. 2 (6):414-27.
- 17. Parent, B. A., X. Wang, and W. Song. 2002. Stability of the B cell antigen receptor modulates its signaling and antigen-targeting functions. Eur. J. Immunol. 32:1839-46.
- 18. Li, C., K. Siemasko, M. R. Clark, and W. Song. 2002. Cooperative interaction of Igalpha and Igbeta

- of the BCR regulates the kinetics and specificity of antigen targeting. Int. Immunol. 14:1179-91.
- 19. Stoddart, A., M. L. Dykstra, B. K. Brown, W. Song, S. K. Pierce, and F. M. Brodsky. 2002. Lipid Rafts Unite Signaling Cascades with Clathrin to Regulate BCR Internalization. Immunity 17:451-62.
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- 31. Martin D.N., Balgley B., Dutta S., Chen J., Cranford J., Kantartzis S., Rudnick P., DeVoe D.L., Lee C. and Baehrecke E.H. (2007) Proteomic analysis of steroid-triggered autophagic programmed cell death in Drosophila. Cell Death and Differentiation 14, 916-923.
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- 3. Hou, H.Y., Heffer, A., Liu, J., Anderson, W.R., Liu, J., Bowler, T. and Pick, L. (2009) Stripy Ftz target genes are coordinately regulated by Ftz-F1. Dev. Biol. 335:442-453.

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Other accomplishments that entailed the use of the Leica SP5 X:

- Eduardo E. Zattara: Winner of 2016 American Microscopical Society Ralph and Mildred Buchsbaum Prize for Excellence in Photomicrography Color division https://www.researchgate.net/publication/296699445 Winner of the 2016 American Microscopical Society Ralph and Mildred Buchsbaum Prize for Excellence in Photomicrography Color division?ev=prf pub
- 2. Venuti, L. S.; Swierzbinski, M.E., & Herberholz, J. (2014, November). *Investigation of fast autoinhibition in the lateral giant circuit of crayfish.* Poster session presented at the meeting of the Society for Neuroscience, Washington D.C.

Publications that entailed the use of the Zeiss AxioObserver (to date):

1. Bastarrachea LJ, Walsh M, Wrenn SP, Tikekar RV. 2017. Enhanced antimicrobial effect of ultrasound by the food colorant Erythrosin B. Food Research International. In press.

Publications that entailed the use of the PerkinElmer spinning Disk (to date):

- 1. Du, L., A. Zhou, A. Patel, M. Rao, K. Anderson, and S. Roy. (2017). Unique patterns of organization and migration of FGF-expressing cells during Drosophila morphogenesis. *Developmental biology*. doi: 10.1016/j.ydbio.2017.05.009
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Publications that entailed the use of the Nikon Eclipse (to date):

1. Du, L., A. Zhou, A. Patel, M. Rao, K. Anderson, and S. Roy. (2017). Unique patterns of organization and migration of FGF-expressing cells during Drosophila morphogenesis. Developmental biology. doi: 10.1016/j.ydbio.2017.05.009

Appendix 5: Example Rates at Other Institutions

Confocal Rates (updated July 2018)

Facility	Instrument	Hourly
Berkeley Biological Imaging Facility	Zeiss LSM 710	\$40
Cornell University Life Sciences Imaging Core	Zeiss LSM 710	\$35
University of Virginia School of Medicine	Zeiss LSM 510	\$42
University of Virginia School of Medicine	LSM880	\$42
Iowa State University Confocal Facility	Leica SP5 X MP	\$33
Northwestern University Biological Imaging	Leica SP5	\$37
Arizona State Imaging Facility	Leica SP5	\$25
Arizona State Imaging Facility	Leica SP8	\$35
Duke University Light Microscopy Core	Leica SP5	\$30
Duke University Light Microscopy Core	Zeiss LSM 780	\$30
Ohio State University	Olympus FV 1000	\$30
Michigan State U Center Advanced Microscopy	Olympus FV 1000	\$25
University of Connecticut	Leica SP8	\$15
UVA Keck Center	Zeiss LSM 510	\$35
UVA Keck Center	Leica SP5X	\$35
Oregon State University	Zeiss LSM 780	\$30
Oregon State University	Zeiss LSM 780	\$26
Yale School of Medicine	Leica SP5	\$50
Texas A&M	Olympus FV 1000	\$42
Rockefeller University	Leica SP8	\$54
Notre Dame	Nikon A1	\$27
Washington University in St. Louis	Nikon A1	\$32
Perlman School of Medicine	Zeiss LSM 710	\$35
Perlman School of Medicine	Leica STED	\$35
University of California, Davis	Olympus FV 1000	\$35
UC Santa Cruz	Leica SP5	\$25
Indiana University	Leica SP5	\$19
Indiana University	Leica SP8	\$21
Oregon Health and Science University	Zeiss LSM 780	\$40
Oregon Health and Science University	Olympus FV 1000	\$33
U of Georgia Biomedical Microscopy Core	Zeiss LSM 710	\$50
University of Illinois at Chicago	Zeiss LSM 710	\$45
University of Illinois at Chicago	Zeiss LSM 710 Meta	\$41
UC San Diego	Olympus FV 1000	\$36
Baylor College of Medicine	LSM780	\$35
Baylor College of Medicine	Leica SP8	\$35
University of Michigan Medical School	Leica SP5 w/2photon	\$55
U of North Carolina Chapel Hill School of Med	Zeiss LSM 710	\$30
University of Chicago	Leica SP8	\$42
University of Chicago	Leica SP5	\$42
Virgina Common Wealth University	Zeiss LSM 710	\$20
Massachusetts Institute of Technology	Olympus FV 1000	\$38
Georgia Tech	Zeiss LSM 710	\$16
NYU Langone Health	Leica SP5	\$55
University of Colorado, Boulder	Nikon A1	\$45
University of Minnesota	LeicaSP5	\$50
University of Minnesota	Nikon A1	\$50
U of Missouri	LeicaSP8	\$30
Yale School of Medicine	Zeiss LSM 710 MP	\$50
Average Hourly Rate	PC199 F9141 / 10 IAIL	\$36
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DeltaVision Rates (updated July 2018)

		Hourly
Facility	Instrument	Rate
Berkeley Biological Imaging Facility	DeltaVision	\$37
Northwestern University Biological Imaging	DeltaVision	\$25
Duke University Light Microscopy Core	DeltaVision Elite	\$17
Rockefeller University	DeltaVision	\$40
Notre Dame	DeltaVision	\$19
Washington University in St. Louis	DeltaVision	\$32
Perlman School of Medicine	DeltaVision	\$20
University of California, Davis	DeltaVision	\$35
Indiana University	DeltaVision	\$16
U of Arizona, Env Health Sciences	DeltaVision	\$25
Oregon Health and Science University	DeltaVision	\$28
U of Georgia Biomedical Microscopy Core	DeltaVision	\$35
University of Arizona Microscopy Alliance	DeltaVision	\$35
University of Arizona Microscopy Alliance	DeltaVision	\$25
UC San Diego	DeltaVision	\$28
University of Michigan Medical School	DeltaVision	\$50
Massachusetts Institute of Technology	DeltaVision	\$38
Average Hourly Rate		\$30

Widefield Fluorescence Microscope Rates (updated July 2018)

Facility	Instrument	Hourly Rate
Cornell University Life Sciences Imaging Core	Olympus Widefield	\$15
University of Washington Keck Facility	Widefield	\$22
UVA Keck Center	Widefield	\$15
Average Hourly Rate		\$17