



Imaging Core Annual Report FY2019

Dr. Charles Delwiche and Amy Beaven

Table of Contents

Executive Summary	1
Mission of the Facility	2
Introduction and History	2
Organization Structure and Governance	3
Personnel	3
Outreach Activities	4
Summary of Imaging Core Usage	5-9
Operating Cost Analysis	9-11
Proposed Rate Schedules	12
Appendix 1: Past Imaging Core Rates	13
Appendix 2: Summary of Changes in IC Instrumentation	14-15
Appendix 3: Imaging Equipment on Campus	16
Appendix 4: Publications	17-27
Appendix 5: Rates at Other Institutions	28-29

Executive Summary

The Imaging Core (IC) plays a key role in research, training, and education in the life sciences at UMD. In FY19 alone, 57 laboratories from 14 departments in 4 colleges (CMNS, AGNR, ENGR, BSOS), plus 2 off-campus users (Weinberg Medical Physics and the Geneva Foundation) used the facility's high-end light microscopes, as did students in the undergraduate course BSCI421 and the FIRE program. During the past year, the director trained 55 new researchers to independently operate the IC's microscopes, bringing the total number of trained users since FY05 to 854. Access to the facility's microscopes and high-quality training has directly resulted in at least 167 publications (see Appendix 4).

In FY19, total income collected via charge-backs from all microscopes exceeded facility expenses by \$25,576. This surplus is due to a number of factors, including: 1) the Advisory Committee directed the IC to purchase a lower-cost (\$9,180 versus \$17,461) service contract on the DeltaVision starting in FY18 (a savings of \$8,281), 3), the director negotiated with Leica to lower the service contract cost by \$2,781, 4) the IC raised hourly rates by \$2 on the confocal and DeltaVision and by \$5 on the AxioObserver and Nikon scopes beginning February 1, 2018 (again, as per the Advisory Committee) and 5) use of the IC's instruments remains just below FY18's historically high level (Figure 1). FY19 marks the second year in a row that the Leica SP5x collected enough income from charge-backs to pay for both the microscope service contract and the (exceedingly expensive) contract on the white light laser.

If rates remain unchanged and microscope use stays at FY19 levels, the IC account balance will slowly increase. However, as the instruments continue to age, we predict expenses will increase beyond current average levels due to wear and tear on parts and instruments that are not covered under service contracts, including all the various heavily-used objective lenses and, on the Deltavision, the light source and sCMOS camera, which are not covered under the less expensive service contract. In addition, if the IC receives funding to purchase a new confocal (see #2 below), the IC will need to purchase a number of peripheral items for the system including network attached storage, a computer table, and an uninterrupted power supply.

Looking to the future, the IC's two confocals (purchased in 2008 and 2009) will soon become obsolete. Meanwhile, the number of new, high-end microscopes on campus has steadily been increasing, in part due to the IC director's involvement in the acquisition, management, and installation of the PerkinElmer spinning disk, ASI diSPIM lightsheet, and JPK AFM systems for the Imaging Incubator. If the IC is to remain competitive, solvent, and self-sustaining, the director suggests we take the following actions:

1. Maintain current rates in FY20, in spite of the surplus in FY19.
2. Obtain funding for a new, state-of-the-art, confocal microscope. The director started this process in May 2017 by submitting an NIH S10 Shared Instrumentation Grant for a Zeiss LSM 880 Airyscan fast laser scanning confocal microscope. The proposal was not funded, but was resubmitted in May 2018. It received a score of 20, with positive reviews including, "all the reviewers had outstanding enthusiasm for this application, with the instrument expected to have high impact on the users' research productivity". It was recommended for funding by the Program Official in March 2019 and is currently pending administrative review.

Implementing some or all of these recommendations will ensure the IC remains a valuable imaging resource for years to come.

Facility Mission

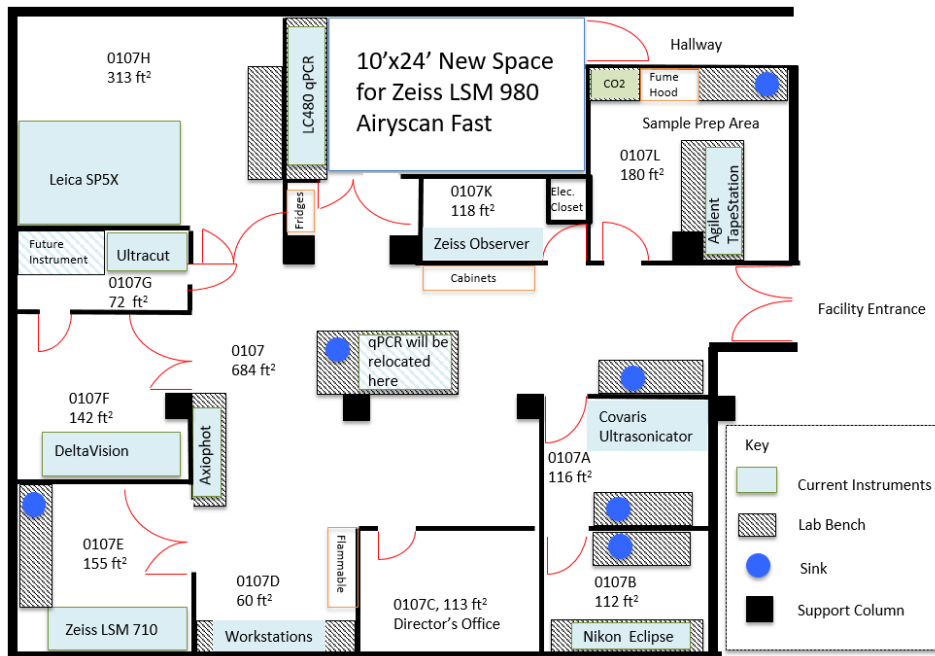
The mission of the Imaging Core (IC) is to enhance research and education by:

1. Providing access to state-of-the-art light microscopy and imaging instrumentation.
2. Offering detailed training opportunities and support in basic and advanced light microscopy techniques.
3. Keeping researchers up to date with the latest technology and innovations in light microscopy.

Introduction/History

The Imaging Core (IC) was established in the year 2000 by the Department of Cell Biology and Molecular Genetics in order to enhance research and education at the University of Maryland. It provides students and faculty members with access to sophisticated light microscopes and imaging instrumentation whose purchase and maintenance costs far exceed the budgets of individual investigators.

The IC facility is located in 0107 Microbiology building, and includes 10 rooms, five of which are dedicated microscope space, office space for the Director, and a wet-bench lab space with fume hood. When first established, the IC contained a single confocal microscope and a deconvolution microscope. Over the years, demand for time on the instruments increased dramatically, necessitating the purchase of a second confocal in 2008. At present, the IC contains 2 laser scanning confocal microscopes (a Zeiss LSM710 and Leica SP5X), a DeltaVision deconvolution/TIRF microscope (installed in March 2014), a Zeiss AxioObserver fluorescence microscope (acquired in February FY14), a Zeiss AxioPhot brightfield microscope, and a Nikon Eclipse upright microscope with color camera (transferred to the facility in October 2016).



In December 2014, a PerkinElmer spinning disk confocal microscope was purchased and installed in the Physical Sciences Complex, and became the first instrument in the new Imaging Incubator facility.

Following that, an ASI diSPIM was installed in April 2017, and a JPK NanoWizard 4a BioAFM in October 2017. The Director of the IC managed these instruments until transferring all responsibilities to the new Director of the IC, Stephan Brenowitz (hired in July 2017). Dr. Brenowitz left the position in April 2019. A search is open for a Lab Manager position to replace him.

The Director of the IC, Amy Beaven, oversees the routine operation of the IC and is available during normal business hours to provide training on all equipment, guidance on experimental design, assistance with image analysis, and technician-assisted microscope operation. Since taking over operation of the IC in November 2005, Ms. Beaven has trained over 799 researchers from at least 13 different departments in six colleges and three different campuses (UMCP, UMB, Shady Grove, Salisbury) of the University of Maryland.

The IC is used by a diverse group of investigators, including undergraduates, graduate students, post-docs, technicians and faculty. In addition, students enrolled in the annual 2-credit class CBMG688W/BSCI427, Principles of Microscopy, gained hands-on experience in the operation of the IC's brightfield, DeltaVision and Leica SP5X confocal microscopes until the fall of 2017, when Professor Stephen Wolniak retired. The course trained an average of fifteen students each year for over eleven years.

In the past, funding for the IC came from a combination of user fees and support from the University of Maryland. In an effort to become self-sustaining, trends in facility income, expenses and instrument usage were analyzed over time (the details of which are published in IC's FY10-18 Annual Reports). The analysis showed that a gradual increase in hourly instrument rates were necessary in order for the facility to become financially independent. As such, user fees were incrementally increased over several years in the hopes that the IC would be able to cover all maintenance and service contract costs through user fees alone. It should be noted that current instrument fees are priced competitively and still below the average rates charged at similar institutions with equivalent instrumentation (see Appendix 5).

Please see Appendix 3 for a complete list of current instrumentation, and Appendix 2 for changes in IC instrumentation since FY05.

Organizational Structure and Governance

- Director of the Facility: Amy Beaven
- Faculty supervisor: Dr. Charles Delwiche, Professor
- Advisory Committee: Kevin McIver, Professor (Chair), Charles Delwiche, Professor (CBMG), Jose Feijo, Professor (CBMG), Antony Jose, Professor (CBMG), Iqbal Hamza, Professor (ANSC), Wolfgang Losert, Professor (PHYS/IPST), Olscan Hollister (CMNS Dean's Office).

Personnel

The Director of the Facility, Amy Beaven, is the only full-time staff member within the facility. She was hired in 2005 to manage the Imaging and Genomics Core facilities and was promoted to Director in 2010. Ms. Beaven received her Master's degree in Biology in 1999 and has over 15 years of intensive experience in imaging techniques, including laser scanning confocal, spinning disk confocal, deconvolution, and TIRF. She is available during the hours of 8am-4:30pm to provide guidance in experimental design, training on all equipment, technician-assisted confocal operation and assistance with image analysis.

Outreach Activities During FY19

1. July, 2019: the director of the IC continued to assist BioENGR/IBBR with their Zeiss LSM 700 system by training students how to use the scope, including proper use of an objective lens inverter.
2. October 2018: the director of the IC organized and hosted a SyGlass virtual reality software demo.
3. October 2018: the director represented the IC by participating in a meeting with the VPR's office to explore Agilent iLab Core Management software.
4. October-December 2018: the IC director completed 7 classes of the LDI Foundations of Leadership and Development.
5. November 2019: the IC took over management of a Covaris Ultra-sonicator, which was relocated to the IC from the IBBR Sequencing Core.
6. January 18, 2019: the IC director participated in an ASI diSPIM alignment session with members of the Losert lab and Drs. Hari Shroff and Harsh Vishwasrao.
7. January 30- February 6, 2019: The IC organized and hosted a Leica SP8 confocal with Lightning super resolution demo.
8. February 2019: the IC director gave members of MRI Global a tour of the IC and participated in a discussion regarding possible collaborations with the campus.
9. February 11-14: The IC organized and hosted a Zeiss AxioScan Z.1 automated slide scanner demo.
10. February-March 2019: the IC director submitted JIT for the NIH S10 Zeiss LSM 880 Airyscan Fast proposal, which was subsequently recommended for funding by the Program Officer. Because NIH would not fund the offline computer workstation, the IC director worked with the Chairs of CBMG and Biology, the assistant Dean of CMNS, and the VPR's office to obtain a Letter of Support for matching funds, which was submitted with the JIT.
11. March 2019: the IC director participated in several troubleshooting sessions with Baltimore Precision Instruments to assist Dr. Stephan Brenowitz (director of the Imaging Incubator) with problems related to the PerkinElmer system.
12. March 2019: the director of the IC trained teaching assistants in BSCI421 (Cell Biology) to use Zeiss AxioObserver and Nikon Eclipse so that they could assist undergraduate students to take images for their laboratory experiments. The director also gave the class a tour of the facility.
13. March 4-14, 2019: The IC organized and hosted a Nikon CSU-W1 spinning disk confocal SoRa superresolution demo.
14. March 22, 2019: the IC director took back over management of the Imaging Incubator from Dr. Stephan Brenowitz.
1. May 31, 2019: The director coordinated with NIH-funded faculty in CBMG, Biology, ANSC, and Physics to resubmit an NIH S10 Shared Instrumentation Grant for the purchase of a new Zeiss LSM 980 Airyscan Fast as a backup plan in case the 2018 proposal is not funded.
15. May 28, 2019: the IC director coordinated installation/training of a new Agilent TapeStation instrument.

Summary of Facility Usage

Starting in FY18, the director began tracking both the billable hours for each instrument, as well as the actual hours each instrument was scheduled/booked and therefore unavailable to other users in order to collect more accurate usage data (Tables 1-2).

In FY19, the Zeiss LSM710 was scheduled an average 27 hours/week and Leica SP5X was used 26 hours/week. The combined average usage of 53 hours/week is a 5% decrease from FY18.

A total of 57 laboratories from 14 departments in 4 colleges (CMNS, AGNR, ENGR, BSOS), plus 2 off-campus users (Weinberg Medical Physics and the Geneva Foundation) used the facility's high-end light microscopes, as did students in the undergraduate course BSCI421 and the FIRE program. During the past year, the director trained 55 new researchers to independently operate the IC's microscopes, bringing the total number of trained users since FY05 to 854.

Table 1: Leica SP5X Summary Data:

Fiscal Year	Income	Total # Hours Charged	Total # Hours Booked	Hours used for UMCP Courses	Total # Training Sessions
2009	\$5,091	346	N/A	0	39
2010	\$18,363	1283	N/A	71	43
2011	\$24,290	1325	N/A	56	35
2012	\$21,882	1021	N/A	63	29
2013	\$21,922	932	N/A	91	35
2014	\$25,161	886	N/A	71	34
2015	\$25,576	1206	N/A	39	27
2016	\$35,865	1602	N/A	36	17
2017	\$31,802	1153	N/A	6	20
2018	\$49,599	1569	1720	0	28
2019	\$39,378	1212	1346	0	9
Total	\$259,552	12534	N/A	433	316

Table 2: Zeiss LSM710 Summary Data:

Fiscal Year	Income	Total # Hours Charged	Total # Hours Booked	Hours used for UMCP Courses	Total # Training Sessions
2010	\$12,370	804	N/A	0	44
2011	\$33,448	1762	N/A	0	33
2012	\$27,895	1244	N/A	0	20
2013	\$33,889	1248	N/A	0	26
2014	\$31,471	1174	N/A	1.5	17
2015	\$27,655	964	N/A	6.5	22
2016	\$32,847	1021	N/A	7	20
2017	\$26,055	688	N/A	29	30
2018	\$35,400	1091	1192	0	13
2019	\$42,551	1280	1398	0	18
Total	\$303,581	11452	N/A	44	243

Table 3: DeltaVision Summary Data:

Fiscal Year	Income	Total # Hours Charged	Total # Hours Booked	Hours used for UMCP Courses	Total # Training Sessions
2014	\$317	32	N/A	0	12
2015	\$6472	376	N/A	24	13
2016	\$6,824	270	N/A	33	20
2017	\$7267	270	N/A	20	21
2018	\$12,674	423	486	0	6
2019	\$8,611	281	338	0	3
Total	\$33,554	1649	N/A	77	75

Table 4: AxioObserver Fluorescence Microscope Data:

Fiscal Year	Income	Total # Hours Charged	Total # Hours Booked	Hours used for UMCP Courses	Total # Training Sessions
2014	\$113	23	N/A	0	0
2015	\$790	155	N/A	0	1
2016	\$1,086	198	N/A	0	5
2017	\$2066	319	N/A	0	22
2018	\$975	117	130	36	8
2019	\$1,338	113	141	57	9
Total	\$5,030	812	N/A	85	45

Table 5: Nikon Eclipse Microscope Data:

Fiscal Year	Income	Total # Hours Charged	Total # Hours Booked	Hours used for UMCP Courses	Total # Training Sessions
2017	\$103	16	N/A	0	1
2018	\$1025	90	101	7	7
2019	\$2045	168	196	9	8
Total	\$1128	106	N/A	16	16

Table 6: Combined Microscope Data by Fiscal Year:

Fiscal Year	Income	Total # Hours Charged	Total # Hours Booked	Hours used for UMCP Courses	Total # Training Sessions
2009	\$5,091	346	N/A	0	39
2010	\$30,733	2,086	N/A	71	87
2011	\$57,739	3,088	N/A	56	68
2012	\$49,778	2,265	N/A	63	49
2013	\$55,811	2,180	N/A	91	61
2014	\$57,061	2,114	N/A	73	63
2015	\$60,175	2,700	N/A	70	63
2016	\$76,622	3,090	N/A	76	62
2017	\$67,194	2,636	N/A	83	93
2018	\$100,126	3290	3630	36	62
2019	\$90,540	2772	3082	0	30
Total	\$560,330	3290	N/A	619	702*

*Including the old Zeiss LSM510, the total # training sessions increases to 854

Figure 1: Total number of hours used on Imaging Core instruments by fiscal year

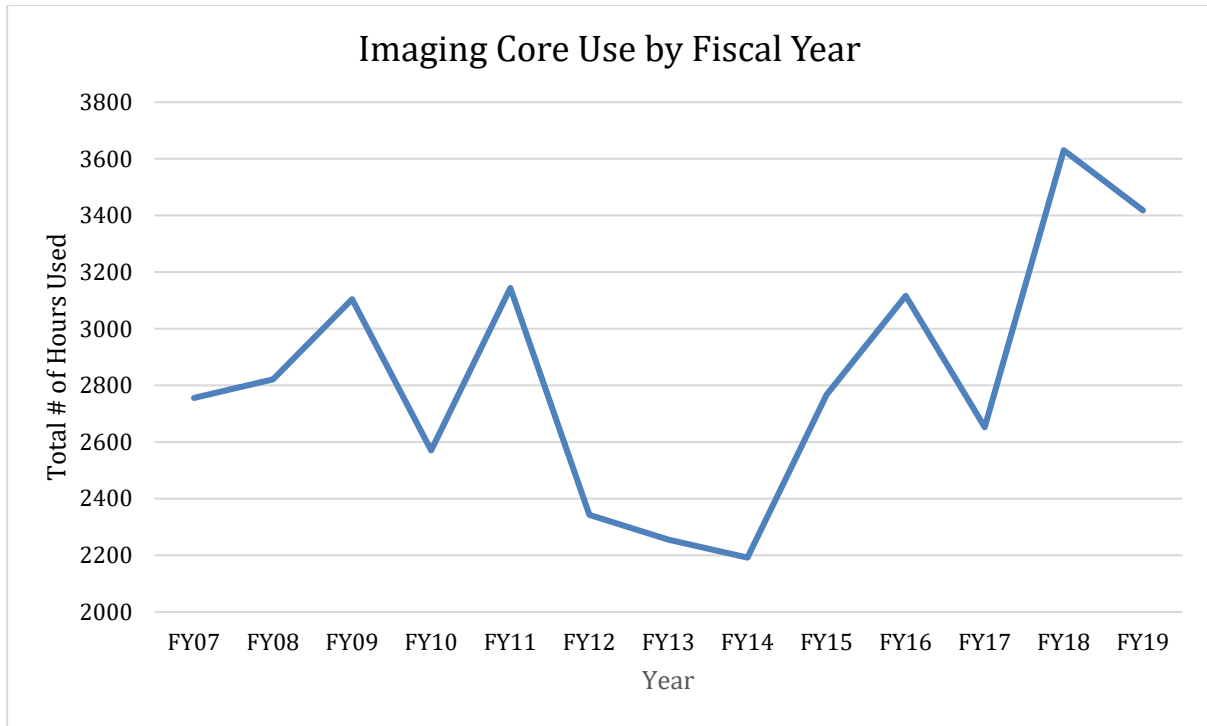


Figure 2: Top Microscope Users by Department FY19

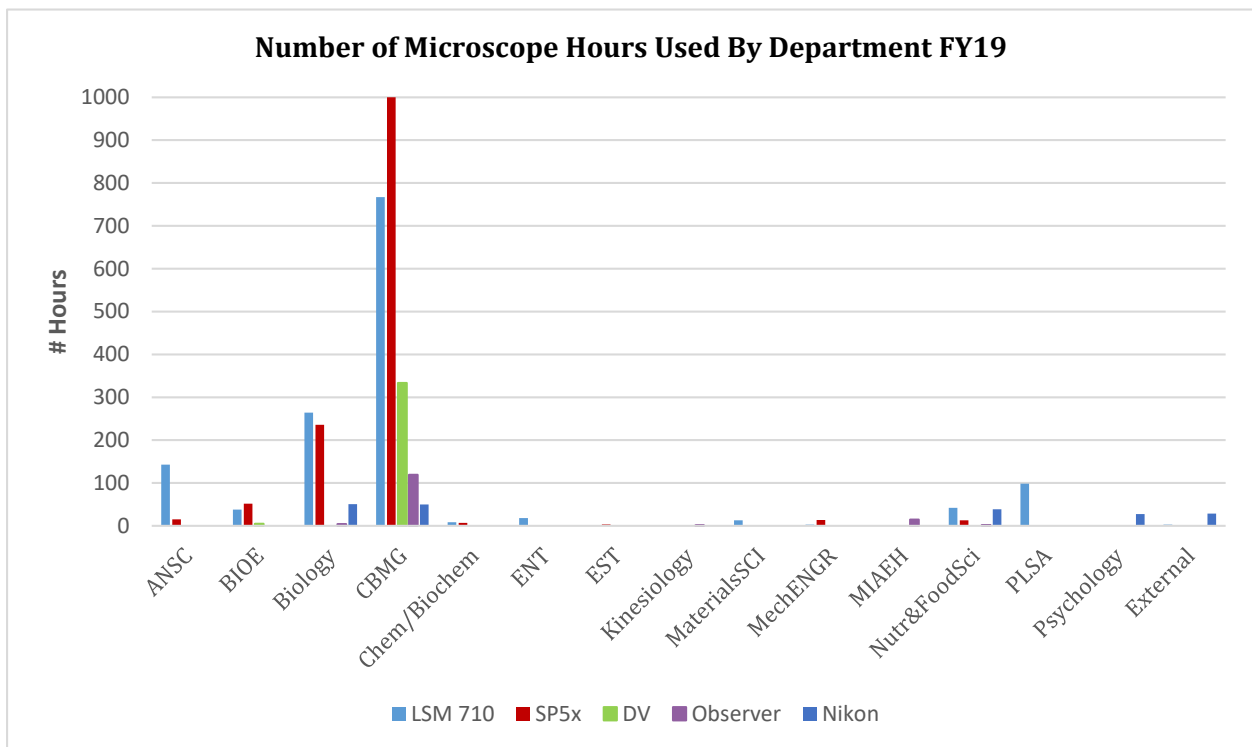


Figure 3: Top Microscope Users FY19

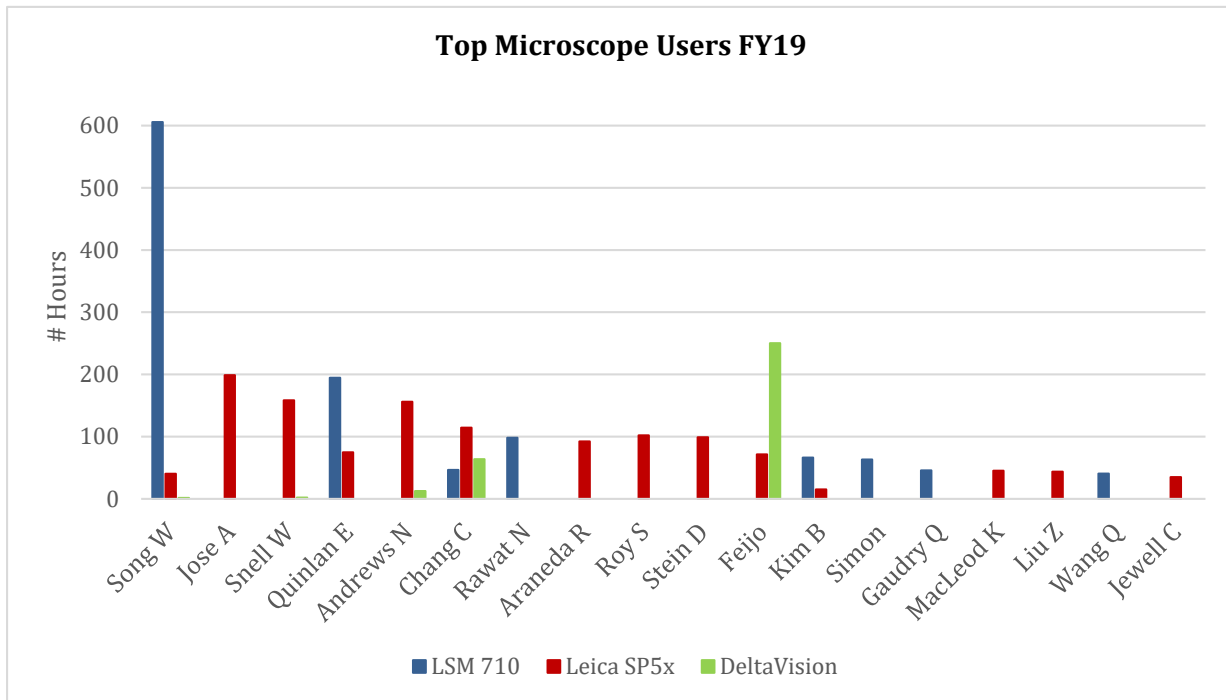


Figure 4: CBMG versus non-CBMG Use of Microscopes FY19

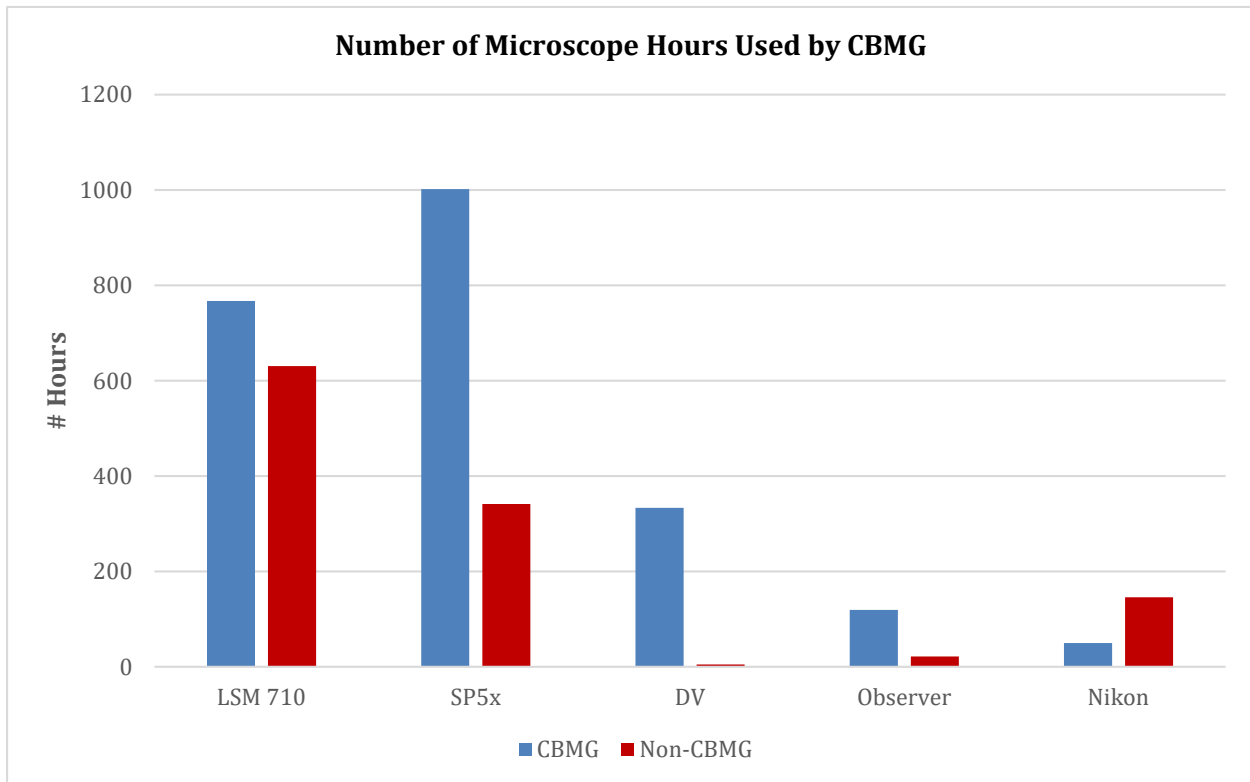
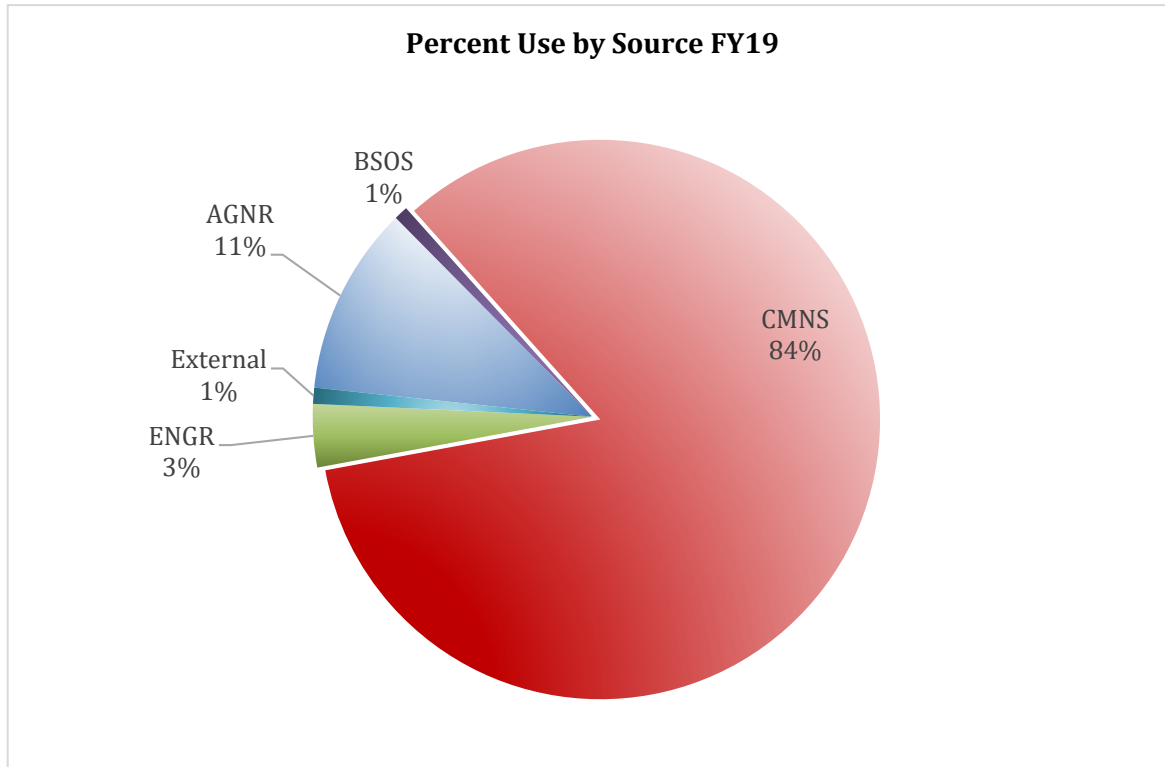


Figure 5: Percent Use by Source FY19



Operating Cost Analysis

In both FY18 and FY19, total income collected via charge-backs exceeded facility expenses. This surplus is due to a number of factors, including: 1) the Advisory Committee directed the IC to purchase a lower-cost (\$9,180 versus \$17,461) service contract on the DeltaVision starting in FY18 (a savings of \$8,281), 2) the director negotiated with Leica to lower the service contract cost by \$2,781, 3) the IC raised hourly rates by \$2 on the confocal and DeltaVision and by \$5 on the AxioObserver and Nikon scopes beginning February 1, 2018 (again, as per the Advisory Committee) and 4) use of the IC's instruments remains just below FY18's historically high level (Figure 1).

Table 7: FY19 Income and Expenses by Microscope

Instrument/ Source Income	Service contract cost	Consumable Cost	Total Expenses	Income	Income - Expenses
Zeiss LSM 710	\$19,969	\$707	\$20,066	\$35,400	\$21,874
Leica SP5 X	\$36,849	\$707	\$39,726	\$49,600	\$1,821
DeltaVision	\$9,450	\$707	\$9,276	\$12,674	(\$1,546)
Zeiss Observer	\$0	\$707	\$96	\$975	\$631
Nikon Eclipse	\$0	\$707	\$96	\$1,025	\$1,338
LSM700 IBBR	N/A	N/A	N/A	\$750	\$750
Total	\$68,491	\$2,828	\$69,097	\$94,673	\$25,576

Table 8: Total Imaging Core Facility Income and Expenses from FY09-FY19

Year	Total Income from Charge-backs*	Total Imaging Core Expenses	Net Balance*
FY2009	\$5,091	\$6,113	(\$1,023)
FY2010	\$30,732	\$29,564	\$1,169
FY2011	\$57,738	\$55,525	\$2,214
FY2012	\$49,778	\$76,562	(\$26,685)
FY2013	\$55,810	\$59,673	(\$3,862)
FY2014	\$57,184	\$59,876	(\$2,692)
FY2015	\$60,303	\$73,955	(\$13,652)
FY2016	\$79,028	\$75,544	\$3,484
FY2017	\$71,458	\$80,234	(\$8,784)
FY2018	\$100,126	\$69,261	\$30,864
FY2019	\$94,673	\$69,097	\$25,967
Total	\$774,423	\$655,412	\$6,510.74

* If instrument income from subsidies associated with the Leica SP5x NSF MRI grant (\$37,500 a year in FY10, FY11, and FY12) is included, the net balance for all instrument at the end of FY19 is \$119,010. The actual account balance in June of 2019 was \$150,660.

Table 9: Cost Breakdown: Leica SP5X

Year	Service Contract Cost	Expenses	Income	Income (subsidies)	Income - Expenses
FY2009	0	\$6,113	\$5,091	0	(\$1,023)
FY2010	\$26,000	\$2,376	\$18,363	\$37,500	\$27,487
FY2011	\$36,075	\$488	\$24,290	\$37,500	\$25,227
FY2012	\$52,296	\$2,056	\$21,882	\$37,500	\$5,032
FY2013	\$37,092	\$1,887	\$21,922	0	(\$17,057)
FY2014	\$37,092	\$360	\$25,160	0	(\$12,290)
FY2015	\$37,743	\$51	\$25,576	0	(\$12,218)
FY2016	\$37,743	\$609	\$35,865	0	(\$2,487)
FY2017	\$39,630	\$616	\$33,158	0	(\$7,088)
FY2018	\$39,630	\$96	\$49,600	0	\$9,873
FY2019	\$36,850	\$707	\$39,378	0	\$1282
Total	\$380,180	\$16,015	\$298,929	\$112,500	\$15,265

Table 10: Cost Breakdown: LSM 710

Year	Service Contract Cost	Expenses	Income	Income (subsidies)	Income - Expenses
FY2009	0	0	0	0	0
FY2010	0	\$1,188	\$12,370	0	\$11,182
FY2011	\$17,730	\$1,232	\$33,448	0	\$14,487
FY2012	\$19,260	\$2,951	\$27,896	0	\$5,685
FY2013	\$19,260	\$1,434	\$33,889	0	\$13,195
FY2014	\$19,260	\$360	\$31,471	0	\$12,211
FY2015	\$19,755	\$51	\$27,655	0	\$7,848
FY2016	\$19,755	\$608	\$32,847	0	\$12,484
FY2017	\$19,969	\$785	\$26,055	0	\$5,301
FY2018	\$19,969	\$96	\$35,400	0	\$15,334
FY2019	\$19,969	\$707	\$42,551	0	\$21,874
Total	\$174,927	\$9,410	\$303,581	0	\$119,603

Table 11: Cost Breakdown: DeltaVision

Year	Service Contract Cost	Expenses	Income	Income (subsidies)	Income - Expenses
FY2014	\$0	\$739	\$371	0	(\$422)
FY2015	\$16,200	\$51	\$6,135	0	(\$10,097)
FY2016	\$16,200	\$323	\$6,824	0	(\$9,700)
FY2017	\$17,461	\$160	\$8,623	0	(\$8,998)
FY2018	\$9,180	\$96	\$12,674	0	\$3,397
FY2019	\$9,450	\$707	\$10,157	0	(\$1,546)
Total	\$68,491	\$2,077	\$41,847	0	(\$28,721)

Projected Cost Analysis:

If rates remain unchanged and microscope use stays at FY19 levels, the IC account balance will continue to grow. This projection does not include potential large one-time expenses, such as objective lens replacements (~\$2000 - \$14,000 depending on the lens), and parts no longer covered under the DeltaVision's more limited service contract (light source and sCMOS camera).

Proposed Rate Schedule:

A thorough examination of confocal microscope rates at other institutions (Appendix 5) shows that the facility’s confocal peak rates are priced slightly lower than the average rate of \$37/hour, and the DeltaVision’s rates are on par with the average. The IC continues to offer highly competitive, low-priced off-peak rates of \$24/hr. The director suggests keeping these rates the same in FY19.

Table 12: Suggested Imaging Core Rates for Academic Year 2019

Instrument	UMD Users	External Users
Leica SP5X, Zeiss LSM710, PerkinElmer Spinning Disk, Hourly Rate for Unassisted Use	\$34/hr peak \$24/off-peak	\$85/hr
DeltaVision Hourly Rate for Unassisted Use	\$30/hr peak \$24/hr off-peak	\$85/hr
Leica SP5X, Zeiss LSM710, DeltaVision, PerkinElmer Spinning Disk 14 hour overnight flat fee, 6pm-8am, M-F	\$150	Inquire
Leica SP5X, Zeiss LSM710, DeltaVision, PerkinElmer Spinning Disk 12 hour day rate (flat fee), 8am-8pm, M-F	\$250	Inquire
Leica SP5X, Zeiss LSM710, DeltaVision, PerkinElmer Spinning Disk 24 hour rate (flat fee), M-F	\$350	Inquire
Leica SP5X, Zeiss LSM710, DeltaVision, PerkinElmer Spinning Disk 24 hour rate (flat fee), Weekend Rate	\$200	Inquire
Leica SP5X, Zeiss LSM710, DeltaVision, PerkinElmer Spinning Disk 48 hour rate (flat fee), Weekend Rate	\$400	Inquire
Leica SP5 X, Zeiss LSM 710, DeltaVision, PerkinElmer Spinning Disk Hourly Rate for Assisted Use	\$50/hr	\$180/hr
Leica SP5X, Zeiss LSM 710, DeltaVision, PE Training Fee (per person)	\$150	\$300
Zeiss AxioObserver, Axiophot and Nikon Eclipse Hourly Rate for Unassisted Use	\$10/hr	\$40/hr
Zeiss AxioObserver, Axiophot and Nikon Eclipse Training Fee (per person)	\$25	\$100

Appendix 1: Past Imaging Core Rates

Equipment	Location	Purchase and/or Installation Date	In-College Rate History (Academic Year)
Zeiss LSM 710 Confocal Microscope	0107E MICB	October 2009 (Purchased by CBMG/CMNS)	2009/2010: \$15.00/hr 2010/2011: \$18.60/hr 2011/2012: \$22.00/hr 2012/2013: \$26.00/hr Peak; \$23.00/hr Off-Peak 2013/2014: \$26.00/hr Peak; \$23.00/hr Off-Peak 2014/2015: \$28.00/hr Peak; \$24.00/hr Off-Peak 2015/2016: \$32.00/hr Peak; \$22.00/hr Off-Peak 2016/2017: \$32.00/hr Peak; \$22.00/hr Off-Peak 2/1/2018: \$34.00/hr Peak; \$24.00/hr Off-Peak 2018/2019: \$34.00/hr Peak; \$24.00/hr Off-Peak
Leica SP5X Confocal Microscope	0107H MICB	December 2008 (Purchased via NSF MRI grant)	2008/2009: \$15.00/hr 2009/2010: \$15.75/hr 2010/2011: \$18.60/hr 2011/2012: \$22.00/hr 2012/2013: \$26.00/hr Peak; \$23.00/hr Off-Peak 2013/2014: \$26.00/hr Peak; \$23.00/hr Off-Peak 2014/2015: \$28.00/hr Peak; \$24.00/hr Off-Peak 2015/2016: \$32.00/hr Peak; \$22.00/hr Off-Peak 2016/2017: \$32.00/hr Peak; \$22.00/hr Off-Peak 2/1/2018: \$34.00/hr Peak; \$24.00/hr Off-Peak 2018/2019: \$34.00/hr Peak; \$24.00/hr Off-Peak
Deltavision Deconvolution/TIRF Microscope	0107F MICB	March 2014 (Purchased by CBMG)	2013/2014: \$10/hr 2014/2015: \$10/hr thru 3/2015 2014/2015: \$28.00/hr Peak; \$24.00/hr Off-Peak 2015/2016: \$28.00/hr Peak; \$22.00/hr Off-Peak 2016/2017: \$28.00/hr Peak; \$22.00/hr Off-Peak 2017/2018: \$30.00/hr Peak; \$24.00/hr Off-Peak 2018/2019: \$34.00/hr Peak; \$24.00/hr Off-Peak
Zeiss AxioObserver Fluorescence	0107K	February 2014 (Relocated from BRB; originally purchased by CBMG)	2013 to 2017: \$5/hr 2/1/18: \$10/hr 2018/2019: \$10/hr
Axiophot Fluorescence Microscope	Main lab	CoolSnap, Elements: July 2007 (purchased by CBMG)	\$2.00/hr since installation
Nikon Eclipse	0107B	October 2017 (relocated)	2016/2017: \$5/hr 2/1/18: \$10/hr 2018/2019: \$10/hr
PerkinElmer confocal spinning disk	B0118 PSC	December 2014 (purchased by Physics for PSC)	2014/2015: \$28.00/hr Peak; \$24.00/hr Off-Peak 2015/2016: \$32.00/hr Peak; \$22.00/hr Off-Peak 2016/2017: \$32.00/hr Peak; \$22.00/hr Off-Peak 2/1/18: \$34.00/hr Peak; \$24.00/hr Off-Peak 2018/2019: \$34.00/hr Peak; \$24.00/hr Off-Peak

Appendix 2: Summary of changes in IC instrumentation since November 2005

- 8/2006: A Mini Med 90 Film Processor (cost: \$3,588.00) replaced the old Konica processor. The department paid \$2,500.00 of the total cost and each of the following PIs contributed \$109: Jonathan Dinman, Jeffrey DeStefano, Kenneth Frauwirth, David Mosser, Anne Simon, Wenxia Song, Richard Stewart and Elizabeth Gantt.
- 10/2006: Dr. Steve Wolniak (Interim Chair of CBMG) procured a Zeiss Axiophot fluorescence microscope for the facility following Dr. Ron Weiner's retirement. A CoolSnap EZ monochrome camera, computer workstation and Nikon Elements software (total cost: \$13,400.00) were purchased in 2007 for the microscope using CBMG funds.
- 4/2007: The 7700 Sequencer Detector was replaced with a Roche LightCycler 480 Real-Time PCR machine, which was purchased by CBMG using the Bioscience Research Building capital equipment funds (and is housed in BRB; see below).
- 8/2007: Due to a drop in usage, the 3100 North DNA sequencer was taken out of operation.
- 12/2008: The instruments in 0107H MICB (two ABI 3100 DNA sequencers, the ABI 3730xl DNA Sequencer and the Roche LightCycler 480 Real-Time PCR machines) were moved to the new Genomics Core, room 2229 Bioscience Research Building.
- 12/2008: The Leica SP5 X confocal microscope was installed in room 0107H MICB. This microscope was obtained by Drs. Ian Mather and Steve Wolniak via an NSF MRI grant.
- 10/2009: The LSM510 confocal microscope was dismantled to make way for the new Zeiss LSM710, which was purchased using college funds, authorized by Dean Allewell.
- 1/2009: Genomics Core Equipment: Bio-Rad CFX 96 Real-time PCR machine was purchased and placed in room 2229 BRB.
- 4/2010: Genomics Core Equipment: Due to a drop in usage, the 3100 "West" DNA sequencer was taken out of operation.
- 7/2011: Genomics Core Equipment: July 2011: Both the 3100 "West" and 3100 "North" DNA sequencers were sold through Terrapin Trader.
- 11/2011: A Thermo Scientific Midi 40 CO2 incubator was purchased using IC funds (\$3,194.00).
- 2/2014: a Zeiss AxioObserver widefield fluorescence microscope was relocated from room 3207 Bioscience Research Building. The microscope was installed in room 0107K and upgraded with a new computer, new software (Zen 2012) and a new power supply, using a combination of departmental and IC funds.
- 3/2014: A DeltaVision Deconvolution/TIRF microscope was installed in room 0107F MICB. The microscope was purchased with departmental and college funds.
- 4/2014: Financial responsibility for the DNA sequencer was transferred to the Biology Dept.
- 8/2014: The Leica SP5 X was upgraded with a new HyD detector and time-gating technology.
- 10/2014: The Zeiss LSM 710 computer was replaced (free upgrade due to computer issues with the old operating system) with a Windows 7 computer and the software was upgraded to the latest version of Zen.
- 12/2014: A PerkinElmer spinning disk confocal microscope was installed in the Satellite Core, room B0118 Physical Sciences Complex.
- 1/2015: As a result of increased Imaging Core responsibilities, management of the Genomics Core was transferred from Amy Beaven to Dr. Yan Wang.
- 7/24/15: The Mini Med 90 Film Processor was taken out of service. It was decontaminated by DES (Cleveland Taylor) and sent to Terrapin Trader.
- 2/2016: The IC implemented a new BSL-2 protocol to allow imaging of approved BSL-2 samples.

- 4/2016: Dr. Charles Delwiche purchased 4 new objective lenses for the DeltaVision and loaned them to the IC: 10x 0.4 NA, 20x 0.75NA, 40x 0.6NA LWD and 40x 1.3NA oil lens.
- 10/2016: A Nikon Eclipse transferred from Dr. Nicole Li's old lab on the 3rd floor of MICB to the IC, room 0107A MICB.
- 4/2017: the ASI diSPIM lighsheet microscope was installed in B0118 PSC on April 4-5 (ahead of the purchase order issued on April 27, 2017).
- 5/2017: The IC acquired room 0105 MICB. The LightCycler 480 qPCR machine was transferred from 2229 BRB to this space, and the Director of the IC now manages the instrument (again).
- 10/7/17: Old DeltaVision Nikon microscope moved to Imaging Incubator in PSC. To be incorporated in the JPK NanoWizard 4a BioAFM.
- 10/16 – 10/18/2017: JPK NanoWizard 4a BioAFM installed (purchase order issued 8/18/17).
- 11/9/2017: Launched new website: <http://biosciencecores.umd.edu/>
- 11/2/2018: The Covaris Ultrasonicator installed in 0107A (transferred from Antony Jose's lab; originally from the IBBR Sequencing Core)
- 5/28/2019: A new Agilent Tapestation was installed in room 0107L MICB.

Appendix 3: Imaging Equipment on Campus

Equipment	Purchase/Installation Date	Owner (purchased by...)	Location	Available to anyone in CBMG?
Zeiss LSM 710 Confocal	10/2009	CBMG/CMNS	CBMG Imaging Core: 0107E MICB	Yes
Leica SP5X Confocal	12/2008	ANSC/CBMG (Purchased via NSF MRI grant)	CBMG Imaging Core: 0107H MICB	Yes
Deltavision Deconvolution/TIRF Microscope	3/2014	CBMG	CBMG Imaging Core: 0107F MICB	Yes
Zeiss AxioObserver Fluorescence	2/2014	CBMG (Relocated from BRB in 2014)	CBMG Imaging Core: 0107K	Yes
Axiophot Brightfield Microscope	7/2007	CBMG (Camera and software)	CBMG Imaging Core: Main lab	Yes
Nikon Eclipse w/color camera	10/2017	CBMG (relocated from Li lab)	CBMG Imaging Core: 0107B	Yes
PerkinElmer confocal spinning disk	12/2014	CMNS (purchased with PSC building funds)	CMNS Imaging Incubator: B0118 PSC	Yes
ASI diSPIM Lightsheet	4/2017	CMNS (purchased w/PSC building funds)	CMNS Imaging Incubator: B0118 PSC	Yes
JPK NanoWizard 4a BioAFM	10/2017	CMNS (purchased with PSC building funds)	CMNS Imaging Incubator: B0118 PSC	Yes
Andor Spinning Disk	August 2016	Sougata Roy (CBMG)	Roy Lab, BRB	No
Zeiss LSM700	~July 2012	IBBR/BioENGR	5131A PLS building	No
Zeiss LSM700 Confocal	Transferred in FY17 from S. Wolniak	Margaret Scull (CBMG)	Scull lab, BRB	No
Zeiss LSM 800 Airyscan Confocal	FY16	Lisa Taneyhill	Taneyhill lab, ANSC	No
Zeiss LSM710	~FY14	Murphy/Peer	Murphy/Peer lab, PLS	No
PerkinElmer Spinning Disk	FY08	Norma Andrews (CBMG)	Andrews lab, BRB	No
Olympus confocal	FY18	BioWorkshop	BioEngineering	Yes

Appendix 4: Publications

Publications that entailed the use of the Zeiss LSM 510:

1. Bish, S. E., W. Song, and D.C. Stein. 2008. Quantification of bacterial invasion into host cells using a beta-lactamase reporter strain: *Neisseria gonorrhoeae* invasion into cervical epithelial cells requires bacterial viability. *Microbes Infect.* 10:1182-1191.
2. Sikes, J. M. & Bely, A. E. Radical modification of the A-P axis and the evolution of asexual reproduction in *Convolutriloba* acoels. *Evolution and Development* 10, 619-631 (2008).
3. The MHC class II-associated invariant chain interacts with the neonatal Fc gamma receptor and modulates its trafficking to endosomal/lysosomal compartments. Ye L, Liu X, Rout SN, Li Z, Yan Y, Lu L, Kamala T, Nanda NK, Song W, Samal SK, Zhu X. *J Immunol.* 2008 Aug 15;181(4):2572-85
4. Activation of the JAK/STAT-1 signaling pathway by IFN-gamma can down-regulate functional expression of the MHC class I-related neonatal Fc receptor for IgG. Liu X, Ye L, Bai Y, Mojidi H, Simister NE, Zhu X. *J Immunol.* 2008 Jul 1;181(1):449-63.
5. Identification and characterization of an alternatively spliced variant of the MHC class I-related porcine neonatal Fc receptor for IgG.
6. Ye L, Tuo W, Liu X, Simister NE, Zhu X. *Dev Comp Immunol.* 2008;32(8):966-79. NF-kappaB signaling regulates functional expression of the MHC class I-related neonatal Fc receptor for IgG via intronic binding sequences. Liu X, Ye L, Christianson GJ, Yang JQ, Roopenian DC, Zhu X. *J Immunol.* 2007 Sep 1;179(5):2999-3011
7. Thyagarajan, R., N. Arunkumar, and W. Song. 2003. Polyvalent antigens stabilize BCR surface signaling microdomains. *J. Immunol.* 170: 6099-106.
8. Onabajo, O., M. Seeley, A. Kale, B. Qualmann, M. Kessels, S-H. Tan, and W. Song. 2008. Mammalian actin-binding protein 1 regulates BCR-mediated antigen processing and presentation in response to BCR activation. *J. Immunol.* 180(10):6685-95.
9. Sharma, S., Orłowski G. and W. Song. 2009. Btk regulates BCR-mediated antigen processing and presentation by controlling the actin cytoskeleton dynamics in B cells. *J. Immunol.* 182: 329-339.
10. Dong CH, Rivarola M, Resnick JS, Maggin BD and Chang C (2008) Subcellular co-localization of Arabidopsis RTE1 and ETR1 supports a regulatory role for RTE1 in ETR1 ethylene signaling. *Plant Journal* 53(2): 275-286
11. Wenming Wang, Alessandra Devoto, John G. Turner, and Shunyuan Xiao. Expression of the Membrane-Associated Resistance Protein RPW8 Enhances Basal Defense Against Biotrophic Pathogens. *Molecular Plant-Microbe Interactions.* 2007 8:966-976
12. Wenming Wang, Xiaohua Yang, Samantha Tangchaiburana, Roland Ndeh, Jonathan E. Markham, Yoseph Tsegaye, Teresa M. Dunn, Guo-Liang Wang, Maria Bellizzi, James F. Parsons, Danielle Morrissey, Janis E. Bravo, Daniel V. Lynch, and Shunyuan Xiao. An Inositolphosphorylceramide Synthase Is Involved in Regulation of Plant Programmed Cell Death Associated with Defense In Arabidopsis. *The Plant Cell* 2008 20:3163-3179
13. Song, W., L. Ma, R. Chen, and D. C. Stein. 2000. Role of lipooligosaccharide in Opa-independent invasion of *Neisseria gonorrhoeae* into human epithelial cells. *J. Exp. Med.* 191 (6):949-60.
14. Cheng, P. C., B. K. Brown, W. Song, and S. K. Pierce. 2001. Translocation of the B cell antigen receptor into lipid rafts reveals a novel step in signaling. *J. Immunol.* 166 (6):3693-701.
15. Song, W. 2001. Signaling, actin dynamics and endocytosis. *Acta Biophysica Sinica.* 17 (1):10-18.
16. Brown, B. K., and W. Song. 2001. The actin cytoskeleton is required for the trafficking of the B cell antigen receptor to the late endosomes. *Traffic.* 2 (6):414-27.
17. Parent, B. A., X. Wang, and W. Song. 2002. Stability of the B cell antigen receptor modulates its signaling and antigen-targeting functions. *Eur. J. Immunol.* 32:1839-46.
18. Li, C., K. Siemasko, M. R. Clark, and W. Song. 2002. Cooperative interaction of Igalpha and Igbeta

- of the BCR regulates the kinetics and specificity of antigen targeting. *Int. Immunol.* 14:1179-91.
19. Stoddart, A., M. L. Dykstra, B. K. Brown, W. Song, S. K. Pierce, and F. M. Brodsky. 2002. Lipid Rafts Unite Signaling Cascades with Clathrin to Regulate BCR Internalization. *Immunity* 17:451-62.
 20. Thompson, M. V., and Wolniak, S. M. 2008. A Plasma Membrane-Anchored Fluorescent Protein Fusion Illuminates Sieve Element Plasma Membranes in Arabidopsis and Tobacco. *Plant Physiology*, 146: 1599-1610
 21. Sirichandra, C., *Gu, D. J., Hu, H.-C., Davanture, M., Lee, S., Djaoui, M., Valot, B., Zivy, M., Leung, J., Merlot, S. and Kwak, J. M. (2009) Phosphorylation of the Arabidopsis AtrbohF NADPH oxidase by OST1 protein kinase. *FEBS Letters* 483 (2009) 2982-2986
 22. Jammes, F., Song, C. J., D. J. Shin, Munemasa, S., Takeda, K., Gu, D., Lee, S., Cho, D. S., Giordo, R., Garg, A., Sritubtim, S., Leonhardt, N., Ellis, E. B., Murata, Y. and Kwak, J. M. (2009) MPK9 and MPK12 are preferentially expressed in guard cells and positively regulate ROS-mediated ABA signaling. will be submitted shortly.
 23. Zhang XN, Mount SM. Two alternatively spliced isoforms of the Arabidopsis thaliana SR45 protein have distinct roles during normal plant development. *Plant Physiol.* 2009 Apr 29 [PubMedID 19403727](#)
 24. Rajagopal, A., Rao, A. U., Amigo, J., Tian, M., Upadhyay, S. K., Hall, C., Uhm, S., Mathew, M. K., Fleming, M. D., Paw, B. H., Krause, M. & Hamza, I. (2008). Haem homeostasis is regulated by the conserved and concerted functions of HRG-1 proteins. *Nature* 453, 1127-31
 25. dos Reis Figueira A, Golem S, Goregaoker SP, Culver JN. A nuclear localization signal and a membrane association domain contribute to the cellular localization of the Tobacco mosaic virus 126-kDa replicase protein. *Virology.* 2002 Sep 15;301(1):81-9. [PubMed PMID: 12359448](#).
 26. Padmanabhan MS, Goregaoker SP, Golem S, Shiferaw H, Culver JN. Interaction of the tobacco mosaic virus replicase protein with the Aux/IAA protein PAP1/IAA26 is associated with disease development. *J Virol.* 2005 Feb;79(4):2549-58. [PubMed PMID: 15681455](#); [PubMed Central PMCID: PMC546588](#).
 27. Padmanabhan MS, Shiferaw H, Culver JN. The Tobacco mosaic virus replicase protein disrupts the localization and function of interacting Aux/IAA proteins. *Mol Plant Microbe Interact.* 2006 Aug;19(8):864-73. [PubMed PMID: 16903352](#).
 28. Dutta S., and Baehrecke E.H. (2008) Warts is required for PI3K-regulated growth arrest, autophagy and autophagic cell death in Drosophila. *Curr. Biol.* 18, 1466-1475.
 29. Juhász G., Hill J.H., Yang Y., Sass M., Baehrecke E.H., Backer J.M. and Neufeld T.P. (2008) The class III PI(3)K Vps34 promotes autophagy and endocytosis but not TOR signaling in Drosophila. *J. Cell Biol.* 181, 655-666.
 30. Berry D.L. and Baehrecke E.H. (2007) Growth arrest and autophagy are required for programmed salivary gland cell degradation in Drosophila. *Cell* 131, 1137-1148.
 31. Martin D.N., Balgley B., Dutta S., Chen J., Cranford J., Kantartzis S., Rudnick P., DeVoe D.L., Lee C. and Baehrecke E.H. (2007) Proteomic analysis of steroid-triggered autophagic programmed cell death in Drosophila. *Cell Death and Differentiation* 14, 916-923.
 32. Martin D. and Baehrecke E.H. (2004) Caspases function in autophagic programmed cell death in Drosophila. *Development* 131, 275-284.
 33. M. Srinivasan and K.A. Frauwirth (2007). "Reciprocal NFAT1 and NFAT2 Nuclear Localization in CD8+ Anergic T Cells is Regulated by Suboptimal Calcium Signaling". *J. Immunol.* 179:3734-3741.
 34. Structural Analysis of "Flexible" Liposomes: Implications for the Transdermal Penetration of Liposomal Structures. Oluwatosin A. Ogunsola, Margaret E. Kraeling, Sheng Zhong, Darrin J. Pochan, Robert L. Bronaugh, Srinivasa R. Raghavan to be submitted to *Langmuir*, June 2009
 35. Kreczko, A., A. Goel, L. Song, and H.-K. Lee (2009) Visual deprivation decreases somatic GAD65 puncta number on layer 2/3 pyramidal neurons in mouse visual cortex. *Neural Plasticity* 2009:

415135.

36. Henry J. Adler, Elena Sanovich, Elizabeth F. Brittan-Powell, Kai Yan, and Robert J. Dooling (2008) WDR1 presence in the songbird basilar papilla. *Hearing Research* 240: 102-111.
37. Isabelle C. Noirot, Henry J. Adler, Charlotte A. Cornil, Nobuhiro Harada, Robert J. Dooling, Jacques Balthazard and Gregory F. Ball (2009) Presence of aromatase and estrogen receptor alpha in the inner ear of zebra finches. *Hearing Research*, 252 (49-55)
38. Title: Evaluation of In Vitro Penetration of Quantum Dot Nanoparticles into Human Skin Authors: M.E.K. Kraeling, O.A. Ogunisola, C.T. Sasik, N.V. Gopee, D.W. Roberts, N.J. Walker, W.W. Yu, V.L. Colvin, P.C. Howard and R.L. Bronaugh. Journal: Manuscript in preparation for *Toxicological Sciences*
39. Padmanabhan MS, Kramer SR, Wang X and Culver JN. 2008. TMV-Aux/IAA interactions: reprogramming the auxin response pathway to enhance virus infection. *J. Virol.* 82:2477-2385.
40. Culver, J.N. and Padmanabhan, M.S. 2007. Virus-induced disease: altering host physiology one interaction at a time. *Annu. Rev. Phytopathol.* 45:221-243.
41. Huang J, Tian L, Peng C, Abdou M, Wen D, Wang Y, Li S, Wang J.(2011) DPP-mediated TGFbeta signaling regulates juvenile hormone biosynthesis by activating the expression of juvenile hormone acid methyltransferase. *Development*, 138(11):2283-91.
42. Chen C, Samuel TK, Krause M, Dailey HA, and Hamza I. Heme utilization in the *Caenorhabditis elegans* hypodermal cells is facilitated by Heme Responsive Gene-2. *J Biol Chem.* 2012;287:9601-12. Epub 2012 Feb 2
43. Chanroj, S. Lu Y, Padmanaban S. Nanatani K, Uozumi N, Rao R, Sze H. Plant-specific cation/H⁺ exchanger 17 and its homologs are endomembrane K⁺ transporters with roles in protein sorting. *J. Biol Chem.* 2011 Sep 30: 286(39):3393-41. Epub 2011 Jul 27.
44. Lu, Y, Chanroj S, Zulkifi L., Johnson MA, Uozumi N, Cheung A, Sze H. Pollen tubes lacking pair of K⁺ transporters fail to target ovules in *Arabidopsis*. *Plant Cell.* 2011 Jan:23 (1)81-93. Epub 2011 Jan 14.
45. Campillo, E, Gaddam S., Mettle-Amuah D., Heneks J. 2012. A tale of two tissues: AtGH9C1 is an endo-B-1,4-glycanase involved in root hair and endosperm development in *Arabidopsis*. *PLOS One* 7(11):e49363.
46. Oluwatosin O., Kraeling M., Zhong S., Pochan J., Bronaugh R and S. Raghavan. 2012. Structural analysis of "flexible" liposome formulations: new insights into the skin-penetrating ability of soft nanostructures. *Soft Matter* (8): 10226-10232.
47. Crivat, G., Lizunov, V., Li, C., Stenkula, K., Zimmergerg, J., Cushman, S., Pick, L. 2013. Insulin stimulates translocation of human GLUT4 to the membrane in fat bodies of transgenic *Drosophila melanogaster*. *PLOS One* 8 (11).
48. Murase S, Lantz CL, Kim E, Gupta N, Higgins R, Stopfer M, ... Quinlan EM. (2016). Matrix Metalloproteinase-9 Regulates Neuronal Circuit Development and Excitability. *Molecular Neurobiology*, 53(5), 3477-93. doi:10.1007/s12035-015-9295-y

Publications that entailed the use of the Leica SP5 X (to date):

1. Sikes, J. M. and Bely, A. E. (2008), Radical modification of the A–P axis and the evolution of asexual reproduction in *Convolutriloba* acoels. *Evolution & Development*, 10: 619–631. doi: 10.1111/j.1525-142X.2008.00276.x
2. Sikes, J.M. and Bely, A.E. (2009), Making heads from tails: Development of a reversed anterior-posterior axis during budding in an acoel. *Devel. Biol.* 338 (1): 86-97.
3. Hou, H.Y., Heffer, A., Liu, J., Anderson, W.R., Liu, J., Bowler, T. and Pick, L. (2009) Stripy Ftz target genes are coordinately regulated by Ftz-F1. *Dev. Biol.* 335:442-453.

4. Zhang, H., Liu, J., Li, C.R., Momen, B., Kohasni, R.A. and Pick, L. (2009). A fly model for diabetes: deletion of *Drosophila* Insulin-Like peptides causes growth defects and metabolic abnormalities. *Proc. Natl. Acad. Sci. U S A.* 106:19617-22.
5. Jammes, F., Song, C. J., D. Shin, Munemasa, S., Takeda, K., Gu, D., Cho, D. S., Lee, S., Giordo, R., Sritubtim, S., Leonhardt, N., Ellis, E. B., Murata, Y. and Kwak, J. M. (2009) Two MAP kinases, MPK9 and MPK12, are preferentially expressed in guard cells and positively regulate ROS-mediated ABA signaling. *Proc. Nat'l. Acad. Sci. USA*, 106: 20520-20525.
6. Kong, D., Cho, D. S., Hu, H.-C., Li, J., Lazzaro, M., Lee, S., Jeon, B.-W., Munemasa, S., Murata, Y., Nam, H. G, Pei, Z.-M. and Kwak, J. M. (2010) Arabidopsis glutamate receptor homologs form Ca²⁺-permeable cation channels and contribute to Ca²⁺ uptake. Submitted.
7. Cho, D.S., Villiers, F., Kroniewicz, L., Lee, S., Zhao, J., Hirschi, K., Leonhardt, N. and Kwak, J. M. (2010) CAX1 and CAX3 contribute to the regulation of cytosolic pH and function in crosstalk between auxin and ABA in guard cells. In preparation.
8. Bely, A.E. and J.M. Sikes (2010). Latent regeneration abilities persist following recent evolutionary loss in asexual annelids. *Proceedings of the National Academy of Sciences* 107:1464-1469.
9. Flannery, A, Czibener, C. and Andrews, N.W. (2010) Palmitoylation-dependent association with CD63 targets the Ca²⁺ sensor synaptotagmin VII to lysosomes. *J. Cell Biol.* 191:599-613.
10. Zattara, E.E. and A.E. Bely (2011). Evolution of a novel developmental trajectory: fission is distinct from regeneration in the annelid *Pristina leidyi*. *Evolution & Development* 13:80-95.
11. Cortez, M., Huynh, C., Fernandes, M.C., Kennedy, K.A., Aderem, A. and Andrews, N.W. (2011) *Leishmania* promotes its own virulence by inducing expression of host CD200. *Cell Host & Microbe* 9:463-471. Highlight in *Nature Reviews Microbiology*
12. Fernandez, M.C., Cortez, M., Flannery, A.R., Tam, C., Mortara, R.A. and N.W. Andrews. (2011) *Trypanosoma cruzi* subverts the sphingomyelinase-mediated plasma membrane repair pathway for cell invasion. *J. Exp Med.* 208(5): 909-21.
13. Nunez-Parra A., Pugh V., Araneda R.C. (2011) Regulation of adult neurogenesis by behavior and age in the accessory olfactory bulb. *Mol Cell Neurosci.* 2011 May 10. [Epub ahead of print]
14. Huang J, Wang Y, Raghavan S, Feng S, Kiesewetter K, Wang J. (2011) Human down syndrome cell adhesion molecules (DSCAMs) are functionally conserved with *Drosophila* Dscam[^{TM1}] isoforms in controlling neurodevelopment. *Insect Biochem Mol Biol.* [Epub ahead of print]
15. Feng S, Huang J, Wang J. (2010) Loss of the Polycomb group gene polyhomeotic induces non-autonomous cell overproliferation. *EMBO Rep.* 12(2):157-63.
16. Clavio PE and KA Frauwirth (2012). Anergic CD8+ lymphocytes have impaired NF-kB activation with defects in p65 phosphorylation and acetylation. *J. Immunol.* 188:1213-1221.
17. Sijacic, P, Wang W and Zhongchi Liu (2011) Recessive antimorphic alleles overcome functionally redundant loci to reveal TS01 function in *Arabidopsis* flowers and meristems. *PLoS Genetics* 8(11): 31002342.
18. Kendall E, Shao C and Don L. DeVoe. Formation of asymmetric folded bilayers by lipid bubble injection in a thermoplastic microfluidic chip. Accepted to *Small*.
19. Shao, C., Kendall E and Don L. DeVoe. Electro-optical BLM chips enabling dynamic imaging of ordered lipid domains. Accepted by *Lab on a Chip*.
20. Chau Huynh, Yuan, X., Miguel, D., Renberg, R., Protchenko, O., Philpott, C., Hamza, I., and Andrews, N. (2012) Heme uptake by *Leishmania amazonensis* is mediated by the transmembrane protein LHR1. *PLoS Pathogens.* 8:7:e1002795.
21. A DEAD box RNA helicase is critical for pre-mRNA splicing, cold-responsive gene regulation, and cold tolerance in *Arabidopsis*. Guan Q, Wu J, Zhang Y, Jiang C, Liu R, Chai C, Zhu J. *Plant Cell.* 2013 25(1):342-56. doi: 10.1105/tpc.112.108340. PMID:23371945

22. A KH domain-containing putative RNA-binding protein is critical for heat stress-responsive gene regulation and thermotolerance in Arabidopsis. Guan Q, Wen C, Zeng H, Zhu J. *Mol Plant*. 2013 6(2):386-95. doi: 10.1093/mp/sss119.PMID:23087326
23. A Bi-functional Xyloglucan Galactosyltransferase is an indispensable salt stress tolerance determinant in Arabidopsis. Li W, Guan Q, Wang ZY, Wang Y, Zhu J. *Mol Plant*. 2013 Jun 22. PMID:23571490
24. Analyzing actin dynamics during the activation of the B cell receptor in live B cells. Liu C, Miller H, Sharma S, Beaven A, Upadhyaya A, Song W. *Biochem Biophys Res Commun*. 2012 Oct 12;427(1):202-6. doi: 10.1016/j.bbrc.2012.09.046. Epub 2012 Sep 17. PMID: 22995298
25. Chunying Kang, Omar Darwish, Aviva Geretz, Rachel Shahan, Nadim Alkharouf, and Zhongchi Liu Genome-Scale Transcriptomic Insights into Early-Stage Fruit Development in Woodland Strawberry *Fragaria vesca*. *Plant Cell* 2013 tpc.113.111732; First Published on June 28, 2013; doi:10.1105/tpc.113.111732
26. Zattara, EE and AE Bely (in press; 2013 expected). Investment regeneration, and asexual reproduction in the annelid *Pristina leidyi*. *Journal of Experimental Zoology Part B: Molecular and Developmental Evolution*.
27. Jeong, J., Lisinski, I, Kadegowda, A.K.G., Shin, H., Xu, J., Wooding, F.B.P., Schaack, J., and Mather, I.H. (2013) A test of current models for the mechanism of milk-lipid secretion, *Traffic*, E-pub June 23, 2013; doi: 10.1111/tra.12087.
28. Geng, C & Paukstelis, P.J. (2014) DNA Crystals as Vehicles for Biocatalysis. *J. Am. Chem. Soc.* 136, 7817-7820.
29. Smith, R. S., Hu H., DeSouza A., Eberly C., Krahe K., Chan W. and Araneda R.C. (2015). Differential muscarinic modulation in the olfactory bulb. *J. Neurosci.* 35 (30): 10773-10785.
30. Ramaswamy B., Kulkarni S. D., Villar P. S., Smith R. S., Eberly C. L., Araneda R.C., Depireux D. A. and B. Shapiro. (2015). Movement of magnetic nanoparticles in brain tissue: neck and isms and safety. *Nanomedicine NMB*.
31. Stein, D. C., LeVan B., Hardy B., Wang L., Zimmerman L., and W. Song (2015). Expression of opacity protein interferes with the transmigration of *Neisseria gonorrhoeae* across polarized epithelial cells. *PlosOne*.
32. Zattara E. and A.E. Bely. (2015). Find taxonomic sampling of nervous systems within Naididae (Annelia: Clitellata) reveals evolutionary lability and revised homologies of annelid neural components. *Frontiers in Zoology*. 12:8.
33. Fernandez M., Corrotte M., Miguel D., Tam C., Andrews N. (2015). The exocyst is required for trypanosome invasion and the repair of mechanical plasma membrane wounds. *Journal of Cell Science*. 128, 27-32.
34. Wu D., Flannery A., Cai H., Ko E., and Cao K.* (2014). Nuclear localization signal deletion mutants of lamin A and progerin reveal insights into lamin A processing and emerin targeting. *Nucleus*, Vol 5-1. doi: 10.4161/nucl.28068. [Highlighted on the cover.]
35. Suresh, R., Chandrasekaran, P., Sutterwala, F. S., & Mosser, D. M. (2016). Complement-mediated 'bystander' damage initiates host NLRP3 inflammasome activation. *Journal Of Cell Science*. 129(9), 1928-1939. doi:10.1242/jcs.179291
36. Xiang J., Forrest I.S., & Pick L. (2015). *Dermestes maculatus*: an intermediate-germ beetle model system for evo-devo. *EvoDevo*. 6, 32. doi:10.1186/s13227-015-0028-0
37. Miller H., Castro-Gomes T., Corrotte M., Tam C., Maugel T.K., Andrews N.W., & Song W. (2015). Lipid raft-dependent plasma membrane repair interferes with the activation of B lymphocytes. *The Journal Of Cell Biology*, 211(6), 1193-205. doi:10.1083/jcb.201505030
38. Sun X, Driscoll MK, Guven C, Das S, Parent CA, Fourkas JT, & Losert W. (2015). Asymmetric nanotopography biases cytoskeletal dynamics and promotes unidirectional cell guidance. *Proceedings Of The National Academy Of Sciences Of The United States Of America*, 112(41), 12557-62. doi:10.1073/pnas.1502970112

39. J. M. Gammon, L. H. Tostanoski, A. R. Adapa, Y. C. Chiu, and C. M. Jewell. (2015). Controlled delivery of a metabolic modulator promotes regulatory T cells and restrains autoimmunity. *J Control Release*. 210, 169-178. doi:10.1016/j.jconrel.2015.05.277.
40. Y. C. Chiu, J. M. Gammon, J. I. Andorko, L. H. Tostanoski, and C. M. Jewell. (2016). Assembly and immunological processing of polyelectrolyte multilayers composed of antigens and adjuvants. *ACS Applied Materials and Interfaces*. (in press)
41. Y. C. Chiu, J. M. Gammon, J. I. Andorko, L. H. Tostanoski, and C. M. Jewell. (2015). Modular Vaccine Design Using Carrier-Free Capsules Assembled from Polyionic Immune Signals. *ACS Biomaterials Science and Engineering*. 1, 1200-1205. doi:10.1021/acsbiomaterials.5b00375. (cover article)
42. P. Zhang, Y. C. Chiu, L. H. Tostanoski, and C. M. Jewell. (2015). Polyelectrolyte Multilayers Assembled Entirely from Immune Signals on Gold Nanoparticle Templates Promote Antigen-Specific T Cell Response. *ACS Nano*. 9, 6465–6477. doi: 10.1021/acsnano.5b02153.
43. Liu C, Richard K, Wiggins M, Zhu X, Conrad DH, & Song W. (2016). CD23 can negatively regulate B-cell receptor signaling. *Scientific Reports*, 6, 25629. doi:10.1038/srep25629
44. Wong, C and Min Wu (2017) Counterfeit Detection Based on Unclonable Feature of Paper Using Mobile Camera *IEEE Transactions on Information Forensics and Security*, VOL. 12, NO. 8, AUGUST 2017.
45. Zheng-Mei Xiong, Mike O'Donovan, Linlin Sun, Ji YoungChoi, Margaret Ren & Kan Cao. (2017). Anti-Aging Potentials of Methylene Blue for Human Skin Longevity. *Scientific Reports*, 7, 2475. doi:10.1038/s41598-017-02419-3
46. B. Duygu Özpolat, Emily S. Sloane, Eduardo E. Zattara, and Alexandra E. Bely. (2016). Plasticity and regeneration of gonads in the annelid *Pristina leidyi*. *EvoDevo*. BioMed Central. doi: 10.1186/s13227-016-0059-1
47. Ronald McNeil and Paul J. Paukstelis. (2017). Core-Shell and Layer-by-Layer Assembly of 3D DNA Crystals. *Advanced Materials*. Doi: 10.1002/adma.201701019
48. Du, L., A. Zhou, A. Patel, M. Rao, K. Anderson, and S. Roy. (2017). Unique patterns of organization and migration of FGF-expressing cells during *Drosophila* morphogenesis. *Developmental biology*. doi: 10.1016/j.ydbio.2017.05.009
49. Zhang, D. & Paukstelis, P.J. Designed DNA Crystal Habit Modifiers, *J. Am. Chem. Soc*, 2017, 139, 1782-1785.
50. McNeil, R. & Paukstelis P.J. Core-shell and Layer-by-layer assembly of 3D DNA Crystals, *Adv. Materials*, 2017, 29
51. Q. Zeng, J. M. Gammon, L. H. Tostanoski, Y. C. Chiu, and C. M. Jewell. "In vivo Expansion of Melanoma-specific T Cells using Microneedle Arrays Coated with Immune-Polyelectrolyte Multilayers" *ACS Biomaterials Science & Engineering* 2017, 5, 195-205.
52. L. H. Tostanoski, Y. C. Chiu, J. M. Gammon, T. Simon, J. I. Andorko, J. S. Bromberg, and C. M. Jewell. "Reprogramming the local lymph node microenvironment promotes tolerance that is systemic and antigen-specific." *Cell Reports* 2016, 16, 2940-2952.
53. Y. C. Chiu, J. M. Gammon, J. I. Andorko, L. H. Tostanoski, and C. M. Jewell. "Assembly and immunological processing of polyelectrolyte multilayers composed of antigens and adjuvants." *ACS Applied Materials and Interfaces* 2016, 8, 18722-18731.
54. Francis NA, Winkowski DE, Sheikhattar A, Armengol K, Babadi B, Kanold PO (2018). Small Networks Encode Decision-Making in Primary Auditory Cortex. *Neuron* 97:885–897.
55. Wang, L.-C.; Litwin, M.; Sahiholnasab, Z.; Song, W.; Stein, D.C. *Neisseria gonorrhoeae* Aggregation Reduces Its Ceftriaxone Susceptibility. *Antibiotics* 2018, 7, 48.
56. Michael M. Wudick, Maria Teresa Portes, Erwan Michard, Paul Rosas-Santiago, Michael A. Lizzio, Custódio Oliveira Nunes, Cláudia Campos, Daniel Santa Cruz Damineli, Joana C. Carvalho, Pedro T. Lima, Omar Pantoja, José A. Feijó. (2018) CORNICHON sorting and regulation of GLR channels

underlie pollen tube Ca²⁺ homeostasis. *Science* 04 May 2018: Vol. 360, Issue 6388, pp. 533-536
DOI: 10.1126/science.aar6464

57. Lerman, MJ, Muramoto, S, Arumugasaamy, N, Van Order, M, Lembong, J, Gerald, AG, Gillen, G, Fisher, JP. 2019. Development of surface functionalization strategies for 3D - printed polystyrene constructs. *J Biomed Mater Res Part B*. 2019: 9999B: 1– 13.
58. Snusha Ravikumar, Sindhuja Devanapally, Antony M Jose. 2019. Gene silencing by double-stranded RNA from *C. elegans* neurons reveals functional mosaicism of RNA interference. *Nucleic Acids Research*. doi: <https://doi.org/10.1101/393074>
59. A.C. Lamont, M.A. Restaino, M.J. Kim, R.D. Sochol, "A Facile Multi-Material Direct Laser Writing Strategy," *Lab on a Chip*, 2019, 19, 2340-2345. DOI: 10.1039/C9LC00398C
60. Sohr A, Du L, Wang R, Lin L, Roy S. Drosophila FGF cleavage is required for efficient intracellular sorting and intercellular dispersal. *J Cell Biol*. 2019 Mar 6;218(5):1653-1669. doi: 10.1083/jcb.201810138. Epub 2019 Feb 26. PMID: 30808704; PMCID: PMC6504889
61. Du L, Sohr A, Yan G, Roy S. Feedback regulation of cytoneme-mediated transport shapes a tissue-specific FGF morphogen gradient. *Elife*. 2018 Oct 17;7. pii:e38137. doi: 10.7554/eLife.38137. PMID:30328809; PMCID:PMC6224196.
62. Du L, Zhou A, Sohr A, Roy S. An Efficient Strategy for Generating Tissue-specific Binary Transcription Systems in Drosophila by Genome Editing. *J Vis Exp*. 2018 Sep 19;(139). doi: 10.3791/58268. PMID: 30295654; PMCID: PMC6235241.

Publications that entailed the use of the LSM710 (to date):

1. Roy V., Smith J.I., Wang J., Stewart J.E., Bentley W.E. & Sintim H.O. (2010) Synthetic analogs tailor native AI-2 signaling across bacterial species. *Journal of the American Chemical Society*, 132 (32), pp 11141–11150
2. Montey KL, Quinlan EM. (2011) Recovery from chronic monocular deprivation following reactivation of thalamocortical plasticity by dark exposure. *Nat Commun*. 2:317.
3. He, K., A. Lee, L. Song, P. O. Kanold, and H.-K. Lee. (2011) AMPA receptor subunit GluR1 (GluA1) serine-845 site is involved in synaptic depression but not in spine shrinkage associated with chemical long-term depression. *Journal of Neurophysiology*, 105: 1897-1907.
4. Dong C.-H., Jang M., Scharein B., Malach A., Rivarola M., Liesch J., Groth G., Hwang I. and Chang C. (2010) Molecular association of the *Arabidopsis* ethylene receptor ETR1 and a regulator of ethylene signaling, RTE1. *J. Biol. Chem*. 285: 40706-40713.
5. Liu, X., Lu, L., Palaniyandi, S., Zeng, R., Gao, L.Y., Mosser, D.M., Roopenian, D.C. and X. Zhu. (2011). The neonatal FcR-mediated presentation of immune-complexed antigen is associated with endosomal and phagosomal pH and antigen stability in macrophages and dendritic cells. *J. Immunol*. 186(8): 4674-86.
6. Meyer, M.T., V. Roy, WE Bentley, and R. Ghodssi. (2011). Development and validation of a microfluidic reactor for biofilm monitoring via optical methods. *J. Micromech. Microeng* (21): 054023 (10pp).
7. Driscoll MK, Albanese J, Xiong ZM, Mailman M, Losert W, and Cao K. (2012). A novel automated image analysis of nuclear shape: What can we learn from a prematurely aged cell? *Aging* 2012 Feb, Vol. 4, No 2
8. Chen C, Samuel TK, Sinclair J, Dailey H and Hamza I. An intercellular heme trafficking protein delivers maternal heme to the embryo during development in *C. elegans*. *Cell*. 2011; 145:720-731
9. Yuan X, Protchenko O, Philpott CC, and Hamza I. Topologically conserved residues direct heme transport in HRG-1-related proteins. *J Biol Chem*. 2012;287:4914-4924. Epub 2011 Dec 15.

10. Recovery from chronic monocular deprivation following reactivation of thalamocortical plasticity by dark exposure. Montey KL, Quinlan EM. *Nature Communications* 2011;2:317
11. HRG1 is essential for heme transport from the phagolysosome of macrophages during erythrophagocytosis. White C, Yuan X, Schmidt PJ, Bresciani E, Samuel TK, Campagna D, Hall C, Bishop K, Calicchio ML, Lapierre A, Ward DM, Liu P, Fleming MD, Hamza I. *Cell Metab.* 2013 Feb 5;17(2):261-70. doi: 10.1016/j.cmet.2013.01.005.
12. Chau Huynh, Yuan, X., Miguel, D., Renberg, R., Protchenko, O., Philpott, C., Hamza, I., and Andrews, N. (2012) Heme uptake by *Leishmania amazonensis* is mediated by the transmembrane protein LHR1. *PLoS Pathogens.* 8:7:e1002795.
13. Ju C, Yoon G.M., Shemansky J.M., Lin D.Y., Ying Z.I., Chang J., Garrett W.M., Kessenbrock M., Groth G., Tucker M.L., Cooper B., Kieber J.J. and Chang C. (2012) CTR1 phosphorylates EIN2 to control ethylene signaling from the ER membrane to the nucleus. *Proc. Natl. Acad. Sci. USA* 109: 19486-19491. (doi: 10.1073/pnas.1214848109).
14. Actin reorganization is required for the formation of polarized B cell receptor signalosomes in response to both soluble and membrane-associated antigens. Liu C, Miller H, Orłowski G, Hang H, Upadhyaya A, Song W. *J Immunol.* 2012 Apr 1;188(7):3237-46. doi: 10.4049/jimmunol.1103065. Epub 2012 Mar 2. PMID: 22387556
15. Levan, A., L. I. Zimmerman, A. C. Mahle, K. V. Swanson, P. Deshong, J. Park, V. L. Edwards, W. Song, and D. C. Stein. 2012. Construction and Characterization of a derivative of *Neisseria gonorrhoeae* strain MS11 devoid of all opa genes. *J. Bacteriol.* 194(23):6468-78.
16. Edwards, V. L., L. C. Wang, V. Dawson, D. C. Stein, and W. Song#. 2013. *Neisseria gonorrhoeae* breaches the apical junction of polarized epithelial cells for transmigration by activating EGFR. *Cell. Microb.* 15(6):1042-57.
17. Arunkumar, N., C. Liu, H. Hang and W. Song#. 2013. Toll-like receptor agonists induce apoptosis in mouse B-cell lymphoma cells by altering NF- κ B activation. *Cell. Mol. Immunol.* In press.
18. K⁺ transporter AtCHX17 with its hydrophilic C tail localizes to membranes of secretory/endocytic system: Role in reproduction and seed set. ChanrojS, Padmanaban S, Czerny DD, Jauh GY, Sze H. *Mol Plant.* 2013 May 23. [Epub ahead of print]
19. Shrivage, B.*, J.H. Hill*, C.Powers, L.Wu and E.Baehrecke. Atg6 is required for multiple vesicle trafficking pathways and hematopoiesis in *Drosophila*. *Development* 140(6): 1321-9 (2013). *These two authors contributed equally.
20. Garg, A. and Wu, L. *Drosophila* Rab14 regulates phagocytosis and antimicrobial peptide expression critical for the immune response to *Staphylococcus aureus*. *Cellular Microbiology*, provisionally accepted.
21. Gupta, A., Terrell, J. L., Fernandes, R., Dowling, M. B., Payne, G. F., Raghavan, S. R. and Bentley, W. E. (2013), Encapsulated fusion protein confers "sense and respond" activity to chitosan-alginate capsules to manipulate bacterial quorum sensing. *Biotechnol. Bioeng.*, 110: 552-562. doi: 10.1002/bit.24711
22. V. Roy, M. T. Meyer, J. A. I. Smith, S. Gamby, H. O. Sintim, R. Ghodssi, and W. E. Bentley, "AI-2 analogs and antibiotics: a synergistic approach to reduce bacterial biofilms," *Applied Microbiology and Biotechnology*, vol. 97, pp. 2627-2638, March 2013.
23. Bloom, S., Williams, A, and MacLeod, K.M. (2014) Heterogeneous calretinin expression in the avian cochlear nucleus angularis. *Journal of the Association for Research on Otolaryngology* 15(4):603-20. doi: 10.1007/s10162-014-0453-0 PMID: 24752525.
24. Gonzalez, E.A., A. Garg, J.Tang, A.Nazario-Toole, L.P.Wu. A Glutamate Dependent Redox System in Blood Cells is Integral for Phagocytosis in *Drosophila melanogaster*. *Current Biology*, 18;23(22): 2319-24 (2013).
25. Xiong, Z., LaDana, C., Wu, D., Cao, K. 2013. An inhibitory role of progerin in the gene induction network of adipocyte differentiation from iPS cells. *Aging* 5 (4) 288-303.

26. Chang J., Clay J.M. and Chang C. (2014) Association of cytochrome *b*₅ with ETR1 ethylene receptor signaling through RTE1 in Arabidopsis. *Plant J.* 77: 558-567. doi: 10.1111/tpj.12401.
27. Seely-Fallen, M., Liu, L., Shapiro, M., Onabojó, O., palaniyandi, S., Zhu, X., Tan, T., Upadhyaya, A., Song, W. (2014). Actin-binding protein 1 links B-cell antigen receptors to negative signaling pathways. *PNAS* 111 (27), p9881-9886.
28. Wu, Chyong-Yi, Hooper, R., Han, K. and Lisa Taneyhill. (2014) Migratory neural crest cell aNcatenin impacts chick trigeminal ganglia formation. *Developmental Biology* (Available online 29 May 2014)
29. Lee, Pei-Chih, Wildt, D. ,Comizzoli, P. (2015) Nucleolar translocation of histone deacetylase 2 is involved in regulation of transcriptional silencing in the cat germinal vesicle. *Biol Reprod.* 115. 129106
30. Shah, A and L. Taneyhill. (2015) differential expression pattern of annexin A6 in chick neural crest and placode cells during cranial gangliogenesis. *Gene Expression Patterns.* 18:21-28.
31. Padmanabhan, R., and L. Taneyhill. (2015). Cadherin-6B undergoes macropinocytosis and clathrin-mediated endocytosis during cranial neural crest cell EMT. *J. Cell. Sci.* 128 (9): 1773-1786.
32. Ju C., Van De Poel B., Cooper E. D. Thierer J. H., Gibbons T. R., Delwiche C. F., Chang C. (2015) conservation of ethylene as a plant hormone over 450 million years of evolution, *Nature Plants* 1:14004.
33. Ketchum, C., Miller H., Song, and Upadhyaya A. 2014. Ligand mobility regulates B cell receptor clustering and signaling activation. *Biophys. J.* 106 (1): 26-36.
34. Zhang H, Xiong Z, and Cao K* (2014). Mechanisms controlling the smooth muscle cell death in progeria via downregulation of Poly(ADP-ribose) polymerase 1. *Proceedings of the National Academy of Sciences.* 111(22):E2261-70. doi: 10.1073/pnas.1320843111. [Highlighted by *UMD Research Roundup*, *UMD-CMNS news*, [San Francisco Chronicle](#) and *Biotrade*.]
35. Srinivasan L., Gurses S.A., Hurley B.E., Miller J.L., Karakousis P.C., Briken V. (2016). Identification of a Transcription Factor That Regulates Host Cell Exit and Virulence of Mycobacterium tuberculosis. *PLoS Pathog.* 12(5): e1005652. doi:10.1371/journal.ppat.1005652
36. Melchiorri, A., Bracaglia, L.G., Kimerer, L.K., Hibino, N., J.P. Fisher, In Vitro Endothelialization of Biodegradable Vascular Grafts via Endothelial Progenitor Cell Seeding and Maturation in a Tubular Perfusion System Bioreactor. *Tissue Eng Part C*, 2016. 22(7). p.663
37. 'M.A. Shumakovich, C.P. Mencio, J.S. Siglin, R.A. Moriarty, H.M. Geller, and K.M. Stroka. (2017). Astrocytes from the brain microenvironment alter migration and morphology of metastatic breast cancer cells, *FASEB Journal*, in press.
38. Wang LC., Yu Q., Edwards V., Lin B., Qiu J., Turner JR., Stein DC., and Song, W. (2017). Neisseria gonorrhoeae infects the human endocervix by activating non-muscle myosin II-mediated epithelial exfoliation. *PLoS Pathogens*, 13(4), e1006269. doi:10.1371/journal.ppat.1006269
39. Gu, Y., Tran, T., Murase, S., Borrell, A., Kirkwood, A., & Quinlan, E. M. (2016). Neuregulin-Dependent Regulation of Fast-Spiking Interneuron Excitability Controls the Timing of the Critical Period. *The Journal of Neuroscience*, 36(40), 10285–10295. doi:10.1523/JNEUROSCI.4242-15.2016
40. Murase S, Lantz CL and Quinlan EM (2017) Light reintroduction following dark exposure rejuvenates the adult visual system via perisynaptic activation of matrix metalloproteinase 9 (MMP-9). *ELife*, Elife. 2017 Sep 6;6. pii: e27345. doi: 10.7554/eLife.27345. PMID: 28875930
41. Bridi MCD, de Pasquale R, Lantz CL, Gu Y, Borrell A, Choi SY, He K, Tran T, Hong SZ, Dykman A, Lee HK, Quinlan EM, Kirkwood A. 2018. Two distinct mechanisms for experience-dependent homeostasis. *Nat Neurosci.* 2018 Jun;21(6):843-850. doi: 10.1038/s41593-018-0150-0. Epub 2018 May 14.

42. Wang, L., Yu, Q., Stein, D. C. and Song, W. (2018). Immunofluorescence Analysis of Human Endocervical Tissue Explants Infected with *Neisseria gonorrhoeae*. *Bio-protocol* 8(3): e2720. DOI: 10.21769/BioProtoc.2720.
43. Lee PC, Wildt DE, Comizzoli P. 2018. Proteomic analysis of germinal vesicles in the domestic cat model reveals candidate nuclear proteins involved in oocyte competence acquisition. *Mol Hum Reprod.* 2018 Jan 1;24(1):14-26. doi: 10.1093/molehr/gax059.
44. Choi, J. Y., Lai, J. K., Xiong, Z., Ren, M., Moorner, M. C., Stains, J. P. and Cao, K. (2018), Diminished canonical β -catenin signaling during osteoblast differentiation contributes to osteopenia in progeria. *J Bone Miner Res.* Accepted Author Manuscript. doi:[10.1002/jbmr.3549](https://doi.org/10.1002/jbmr.3549)
45. Chen, N., Zhang, J., Mei, L., and Wang, Q. (2018). Ionic Strength and pH Responsive Permeability of Soy Glycinin Microcapsules. *Langmuir* 34 (33), 9711-9718. DOI: 10.1021/acs.langmuir.8b01559
46. Zhang X, Coates K, Dacks A, Gunay C, Lauritzen S, Li F, Calle-Schuller S, Bock D, Gaudry Q. Local synaptic inputs support opposing, network-specific odor representations in a widely projecting modulatory neuron. *eLife* 2019;8:e46839
47. Zhang X, Coates K, Dacks A, Gunay C, Lauritzen S, Li F, Calle-Schuller S, Bock D, Gaudry Q. Local synaptic inputs support opposing, network-specific odor representations in a widely projecting modulatory neuron. *eLife* 2019;8:e46839

Publications that entailed the use of the DeltaVision (to date):

1. Renberg R, Yuan X, Samuel T, Miguel D, Hamza I, Andrews N, Flannery A. (2015) The heme transport capacity of LHR1 determines the extent of virulence in *Leishmania amazonensis*. *PlosOne*.
2. Bai, X., Zhang, Y., Huang, L., Wang, J., Li, W., Niu, L., ... Liu, C. (2016). The early activation of memory B cells from Wiskott-Aldrich syndrome patients is suppressed by CD19 down regulation. *Blood*. doi:10.1182/blood-2016-03-703579
3. Ortiz-Ramírez C, Michard E, Simon AA, Damineli, D, Hernández-Coronado M, Becker JD¹ and Feijó JA (2017) Glutamate Receptor-like channels are essential for chemotaxis and reproduction in mosses. *Nature*, 10.1038/nature23478
4. Damineli DSC, Portes MT, Feijó JA (2017) Oscillatory signatures underlie growth regimes in Arabidopsis pollen tubes: a computational package to estimate tip location, periodicity and synchronization in live cells. *J. Exp. Bot.* doi:10.1093/jxb/erx032^[1]_{SEP}
5. Michard E, Simon AA, Tavares B, Wudick MM, Feijó JA (2016) Signaling with ions: the keystone for apical cell growth and morphogenesis in pollen tubes. *Plant Physiol.* 173:91-111
6. Laranjeira-Silva, MF, Wang, W, Samuel, TK, Maeda, FY, Michailowsky, V, Hamza, I, Liu, Z and Andrews, NW. A MFS-type plasma membrane transporter required for *Leishmania* virulence protects the parasites from iron toxicity. *PLoS Pathogens*, 14(6):e10077140. doi: 10.1371/journal.ppat.1007140, 2018.
7. Michailowsky, V., Li, H., Mitra, B., Iyer, S.R., Mazála, D.A.G., Corrotte, M., Wang, Y., Chin, E.R., Lovering, R.M. and Andrews, N.W. Defects in sarcolemma repair and skeletal muscle function after injury in a mouse model of Niemann-Pick Type A/B disease. *Skeletal Muscle*, 9(1):1. doi: 10.1186/s13395-018-0187-5, 2019.

Other accomplishments that entailed the use of the Leica SP5 X:

1. Eduardo E. Zattara: Winner of 2016 American Microscopical Society Ralph and Mildred Buchsbaum Prize for Excellence in Photomicrography Color division
<https://www.researchgate.net/publication/296699445> Winner of the 2016 American Micros

[cological Society Ralph and Mildred Buchsbaum Prize for Excellence in Photomicrography Color division?ev=prf pub](#)

2. Venuti, L. S.; Swierzbinski, M.E., & Herberholz, J. (2014, November). *Investigation of fast autoinhibition in the lateral giant circuit of crayfish*. Poster session presented at the meeting of the Society for Neuroscience, Washington D.C.

Publications that entailed the use of the Zeiss AxioObserver (to date):

1. Bastarrachea LJ, Walsh M, Wrenn SP, Tikekar RV. 2017. Enhanced antimicrobial effect of ultrasound by the food colorant Erythrosin B. Food Research International. In press.

Publications that entailed the use of the PerkinElmer spinning Disk (to date):

1. Du, L., A. Zhou, A. Patel, M. Rao, K. Anderson, and S. Roy. (2017). Unique patterns of organization and migration of FGF-expressing cells during Drosophila morphogenesis. *Developmental biology*. doi: 10.1016/j.ydbio.2017.05.009
2. M.T. Doolin and K.M. Stroka. (2018) Physical confinement alters cytoskeletal contributions towards human mesenchymal stem cell migration, *Cytoskeleton*, 75(3): 103-117.
3. M.A. Pranda*, K.M. Gray*, G.M. Dawson, A.J.L. DeCastro, J.W. Jung, and K.M. Stroka#. Tumor cell mechanosensing during incorporation into the brain microvascular endothelium, *Cellular and Molecular Bioengineering* (accepted, 2019). (*, equal contribution). *** invited as part of Cellular and Molecular Bioengineering Young Innovator Award special issue. *Accepted but not yet published as of 8/1/19/

Publications that entailed the use of the Nikon Eclipse (to date):

1. Du, L., A. Zhou, A. Patel, M. Rao, K. Anderson, and S. Roy. (2017). Unique patterns of organization and migration of FGF-expressing cells during Drosophila morphogenesis. *Developmental biology*. doi: 10.1016/j.ydbio.2017.05.009

Appendix 5: Example Rates at Other Institutions

Confocal Rates (updated July 2019)

Facility	Instrument	Hourly Rate
Berkeley Biological Imaging Facility	Zeiss LSM 710	\$ 40.00
Cornell University Life Sciences Imaging	Zeiss LSM 710	\$ 35.00
University of Virginia School of Medicine	LSM880	\$ 45.00
Northwestern University Biological	Leica SP5	\$ 39.00
Northwestern University Biological	Zeiss LSM 510	\$ 40.00
Arizona State Imaging Facility	Leica SP5	\$ 25.00
Arizona State Imaging Facility	Leica SP8	\$ 35.00
Duke University Light Microscopy Core	Leica SP5	\$ 30.00
Duke University Light Microscopy Core	Zeiss LSM 780	\$ 30.00
Michigan State U Center Advanced	Zeiss LSM 510	\$ 25.00
Michigan State U Center Advanced	Olympus FV 1000	\$ 25.00
University of Washington Keck Facility	Zeiss LSM 510	\$ 45.00
University of Connecticut	Leica SP8	\$ 15.00
UVA Keck Center	Zeiss LSM 510	\$ 35.00
UVA Keck Center	Leica SP5X	\$ 35.00
Oregon State University	Zeiss LSM 780	\$ 30.00
Oregon State University	Zeiss LSM 780	\$ 26.00
Yale School of Medicine	Leica SP5	\$ 50.00
Texas A&M	Olympus FV 1000	\$ 42.00
Boise State University	Zeiss LSM 510	\$ 19.59
Rockefeller University	Leica SP8	\$ 56.00
Notre Dame	Nikon A1	\$ 31.00
Washington University in St. Louis	Nikon A1	\$ 32.00
Perlman School of Medicine	Zeiss LSM 710	\$ 42.00
University of California, Davis	Olympus FV 1000	\$ 35.00
UC Santa Cruz	Leica SP5	\$ 25.00
Indiana University	Leica SP5	\$ 19.75
Indiana University	Leica SP8	\$ 21.75
U of Georgia Biomedical Microscopy Core	Zeiss LSM 710	\$ 50.00
University of Illinois at Chicago	Zeiss LSM 710	\$ 45.00
University of Illinois at Chicago	Zeiss LSM 710 Meta	\$ 41.00
University of Illinois Urbana-Champaign	Zeiss LSM 710	\$ 35.00
University of Illinois Urbana-Champaign	Zeiss LSM 880	\$ 44.00
UC San Diego	Olympus FV 1000	\$ 36.00
Baylor College of Medicine	LSM880	\$ 35.00
Baylor College of Medicine	LSM780 Fast	\$ 35.00
Baylor College of Medicine	Leica SP8	\$ 35.00
University of North Carolina Chapel Hill	Zeiss LSM 710	\$ 35.90
University of Chicago	Leica SP8	\$ 42.44
University of Chicago	Leica SP5	\$ 42.44
Virginia Commonwealth University	Zeiss LSM 710	\$ 20.00
Massachusetts Institute of Technology	Olympus FV 1000	\$ 40.00
Georgia Tech	Zeiss LSM 710	\$ 16.00
NYU Langone Health	Leica SP5	\$ 55.00
NYU Langone Health	Zeiss LSM 880	\$ 65.00
University of Minnesota	LeicaSP5	\$ 52.50
University of Minnesota	Nikon A1	\$ 52.50
U of Missouri	LeicaSP8	\$ 35.00
Albert Einstein College of Medicine	LeicaSP5	\$ 43.00
Albert Einstein College of Medicine	LeicaSP8	\$ 43.00
Average Hourly Rate		\$ 37.00

DeltaVision Rates (updated July 2019)

Facility	Instrument	Hourly Rate
Berkeley Biological Imaging Facility	DeltaVision	\$ 37.00
Northwestern University Biological Imaging	DeltaVision	\$ 25.00
Duke University Light Microscopy Core	DeltaVision Elite	\$ 17.00
Rockefeller University	DeltaVision	\$ 42.00
Notre Dame	DeltaVision	\$ 19.00
University of California, Davis	DeltaVision	\$ 35.00
Indiana University	DeltaVision	\$ 16.50
U of Georgia Biomedical Microscopy Core	DeltaVision	\$ 35.00
University of Arizona Microscopy Alliance	DeltaVision	\$ 35.00
University of Arizona Microscopy Alliance	DeltaVision	\$ 25.00
UC San Diego	DeltaVision	\$ 28.00
University of Michigan Medical School	DeltaVision	\$ 20.00
Massachusetts Institute of Technology	DeltaVision	\$ 40.00
Koch Institute at MIT	DeltaVision	\$ 40.00
Average Hourly Rate		\$ 30.00

Widefield Fluorescence Microscope Rates (updated July 2019)

Facility	Instrument	Hourly Rate
Cornell University Life Sciences Imaging Core	Olympus Widefield	\$ 15.00
University of Washington Keck Facility	Widefield	\$ 22.00
UVA Keck Center	Widefield	\$ 15.00
UC Santa Cruz	Zeiss Axioskop	\$ 8.00
University of Illinois Urbana-Champaign	Zeiss Widefield	\$ 22.00
Perlman School of Medicine	Zeiss Observer	\$ 20.00
Average Hourly Rate		\$ 17.00